



## INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

(51) International Patent Classification 6 : C12N 15/53, 15/82, C12Q 1/26, 1/02, A01H 5/00		A1	(11) International Publication Number: WO 97/49816
			(43) International Publication Date: 31 December 1997 (31.12.97)
(21) International Application Number: PCT/US97/11295		(81) Designated States: AL, AM, AU, AZ, BA, BB, BG, BR, BY, CA, CN, CU, CZ, EE, GE, HU, IL, IS, JP, KG, KP, KR, KZ, LC, LK, LR, LT, LV, MD, MG, MK, MN, MX, NO, NZ, PL, RO, RU, SG, SI, SK, SL, TJ, TM, TR, TT, UA, US, UZ, VN, YU, ARIPO patent (GH, KE, LS, MW, SD, SZ, UG, ZW), Eurasian patent (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM), European patent (AT, BE, CH, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE), OAPI patent (BF, BJ, CF, CG, CI, CM, GA, GN, ML, MR, NE, SN, TD, TG).	
(22) International Filing Date: 26 June 1997 (26.06.97)		<p>Published With international search report.</p>	
(30) Priority Data: 60/021,364 27 June 1996 (27.06.96) US			
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(54) Title: PLANT GENE FOR <i>p</i> -HYDROXYPHENYLPYRUVATE DIOXYGENASE			
<p>1 CAAGAAACGNGTCGNGCAGCTGCTCAGCGATGATCAGATCAAGGAGTGTGAGGAATTAGG.</p> <p>61 GATTCTTNTAGACAGAGATGATCAAGGGACGTTNCTTCAAATCTNCACAAAACCACTAGG</p> <p>121 TGACAGGCCGACGNTATTTATAGAGATAATCCAGAGNGTAGGATGCATGATGAAAGATGT</p> <p>181 GGAAGGGGANGGCTTACCAGAGTGGAGNATNTNGTGGTTTTGGCAAAGGCAATT</p>			
(57) Abstract			
<p>The invention relates to the isolation and modification of nucleic acid sequences encoding <i>p</i>-hydroxyphenylpyruvate dioxygenase enzyme from plants. These nucleic acid sequences were used to establish methods of identification of new herbicidal compounds that inhibit the activity of this enzyme, and to prepare new crop plants that are tolerant to the herbicidal action of inhibitors of this enzyme. Chimeric genes comprising nucleic acid fragments containing all or part of the nucleic acid sequences encoding <i>p</i>-hydroxyphenylpyruvate dioxygenase may be used to produce active plant <i>p</i>-hydroxyphenylpyruvate dioxygenase enzyme in microorganisms, and to cause the production of modified forms of the enzyme in plants that may render such plants tolerant to inhibitors of the enzyme.</p>			

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TITLEPLANT GENE FOR *p*-HYDROXYPHENYLPYRUVATE DIOXYGENASEFIELD OF THE INVENTION

This invention relates to the isolation and modification of nucleic acid  
5 encoding *p*-hydroxyphenylpyruvate dioxygenase enzyme from plants. These  
nucleic acid sequences were used to establish methods of identification of new  
herbicidal compounds that inhibit the activity of this enzyme, and to prepare new  
crop plants that are tolerant to the herbicidal action of inhibitors this enzyme.  
Chimeric genes comprising nucleic acid fragments containing all or part of the  
10 nucleic acid sequences encoding *p*-hydroxyphenylpyruvate dioxygenase may be  
used to produce active plant *p*-hydroxyphenylpyruvate dioxygenase enzyme in  
microorganisms, and to cause the production of modified forms of the enzyme in  
plants that may render such plants tolerant to inhibitors of the enzyme.

BACKGROUND OF THE INVENTION

15 Bleaching herbicides affect plant chloroplasts by decreasing their  
chlorophyll and carotenoid content. Several bleaching herbicides are known to  
inhibit the enzyme phytoene desaturase, resulting in the accumulation of phytoene  
in treated plants. However, compounds of the benzoyl cyclohexane-1,3-dione  
type cause the accumulation of phytoene in plants but are not inhibitors of  
20 phytoene desaturase *in vitro* (Sandmann, G., et al. (1990) *Pestic. Sci.* 30:353-355).  
Subsequent work revealed that these compounds are effective inhibitors of  
*p*-hydroxyphenylpyruvate dioxygenase (*p*-hydroxyphenylpyruvate:oxygen  
oxidoreductase EC 1.13.11.27), a key enzyme in the biosynthesis of  
plastoquinones and tocopherols (Schulz, A., et al. (1993) *FEBS Lett.*  
25 318:162-166). Based on the observation that phytoene desaturase requires a  
quinone as an electron acceptor, these authors postulated that by inhibiting  
*p*-hydroxyphenylpyruvate dioxygenase, these herbicides act indirectly on  
phytoene desaturase by blocking the biosynthesis of quinones.

The proposal that *p*-hydroxyphenylpyruvate dioxygenase is essential for  
30 carotenoid biosynthesis has received support from genetic studies in the plant  
model system *Arabidopsis thaliana*. Mutations in the *pds1* and *pds2* genetic loci  
result in mutant plants that accumulate phytoene. However, genetic mapping of  
these mutant genes indicates that they do not correspond to the gene encoding the  
enzyme phytoene desaturase. The *pds1* mutation can be rescued by homogentisic  
35 acid, the substrate of *p*-hydroxyphenylpyruvate dioxygenase. Therefore, this  
mutation corresponds to a defect in the activity of *p*-hydroxyphenylpyruvate  
dioxygenase (Norris, S. R., et al. (1995) *Plant Cell* 7:2139-2149).

In light of these disclosures, *p*-hydroxyphenylpyruvate dioxygenase is a promising new target for new herbicidal compounds. Research aimed at discovering new herbicides based on this mode of action would be greatly facilitated by the isolation of the plant gene encoding this enzyme and by the functional expression of this gene in transgenic organisms. For example, active enzyme produced in recombinant microorganisms could be used to establish screening methods for the identification of novel active compounds and to obtain structural and mechanistic information useful to guide further chemical synthesis. Furthermore, isolation of this gene would facilitate research aimed at generating mutant, herbicide-tolerant versions of the enzyme that may confer herbicide resistance to transgenic plants.

A partial sequence of an *Arabidopsis thaliana* cDNA with homology to corresponding mammalian sequences encoding *p*-hydroxyphenylpyruvate dioxygenase has been identified (GenBank Accession No. T20952), but this truncated sequence is insufficient to identify an active plant *p*-hydroxyphenylpyruvate dioxygenase. WO 96/38567 A2 addresses the utility that would be attached to a DNA sequence of a *p*-hydroxyphenylpyruvate dioxygenase gene, but there is no biochemical evidence of function associated with the sequences disclosed.

#### SUMMARY OF THE INVENTION

This invention pertains to the isolation and characterization of nucleic acid fragments encoding plant *p*-hydroxyphenylpyruvate dioxygenase enzymes. More specifically, this invention pertains to isolated nucleic acid fragments encoding the *p*-hydroxyphenylpyruvate dioxygenase enzymes from *Arabidopsis thaliana* and *Zea mays*.

This invention also pertains to the production of active plant *p*-hydroxyphenylpyruvate dioxygenase enzyme in *E. coli*. In one embodiment, a chimeric gene comprising a nucleic acid fragment encoding a polypeptide that possesses *p*-hydroxyphenylpyruvate dioxygenase activity, operably linked to regulatory sequences that direct gene expression in *E. coli*, is claimed. In another embodiment, a plasmid vector comprising said chimeric gene is disclosed. In yet another embodiment, a transformed *E. coli* comprising a chimeric gene consisting of a nucleic acid fragment encoding a polypeptide that possesses *p*-hydroxyphenylpyruvate dioxygenase activity is disclosed.

This invention also pertains to a method of identifying substances that inhibit the rate of the reaction of *p*-hydroxyphenylpyruvate dioxygenase enzyme. In one embodiment, the invention pertains to an assay for the detection of inhibitors of *p*-hydroxyphenylpyruvate dioxygenase wherein a polypeptide

derived from a transformed *E. coli* that displays *p*-hydroxyphenylpyruvate dioxygenase activity is incubated in the presence of a test substance. Following incubation, *p*-hydroxyphenylpyruvate dioxygenase enzymatic activity is measured wherein a reduction of enzymatic activity is indicative of the inhibitory capacity of the test substance. Enzymatic activity can be measured by any appropriate means, including but not limited to oxygen utilization, carbon dioxide release, homogentisate production, and loss of *p*-hydroxyphenylpyruvate. Results are quantified by radiometric, colorimetric or chromatographic means.

In another embodiment, this invention pertains to plants that are substantially tolerant to the application of at least one compound that inhibits the rate of the reaction of *p*-hydroxyphenylpyruvate dioxygenase. Plants may be rendered tolerant by overexpression of the wild-type *p*-hydroxyphenylpyruvate dioxygenase, by expression of a naturally-occurring resistant variant of this enzyme, or by expression of an altered form of *p*-hydroxyphenylpyruvate dioxygenase that is resistant to the action of compounds that are inhibitory to the wild-type enzyme.

A further embodiment of the invention is an isolated nucleic acid fragment comprising a member selected from the group consisting of:

- (a) an isolated nucleic acid fragment as set forth in SEQ ID NO:16;
- (b) an isolated nucleic acid fragment that is essentially similar to an isolated nucleic acid fragment as set forth in SEQ ID NO:16; and
- (c) an isolated nucleic acid fragment that is complementary to (a) or (b).

#### BRIEF DESCRIPTION OF THE DRAWINGS AND SEQUENCE DESCRIPTIONS

The invention can be more fully understood from the following detailed description and the accompanying drawings and the sequence descriptions which form a part of this application.

Figure 1 presents a partial nucleic acid sequence of an expressed sequence tag (EST) bearing GenBank Accession No. T92052 obtained from an *Arabidopsis thaliana* cDNA library. This sequence was contained in clone 91B13T7 of the library.

Figure 2 presents the nucleic acid sequence of the cloned cDNA encoding a full-length form of *Arabidopsis thaliana p*-hydroxyphenylpyruvate dioxygenase enzyme, as it was initially determined (SEQ ID NO:2). Translation start and stop codons are underlined. Selected restriction sites are indicated.

Figure 3 presents the amino acid sequence comparison between full-length *p*-hydroxyphenylpyruvate dioxygenases from *Arabidopsis thaliana* (SEQ ID NO:15) and *Zea mays* (SEQ ID NO:11) and the *p*-hydroxyphenylpyruvate dioxygenase enzymes derived from human (SEQ ID NO:6, GenBank Acc.

5 No. U29895), pig (SEQ ID NO:7, GenBank Acc. No. D13390), mouse (SEQ ID NO:8, GenBank Acc. No. D29987) and rat (SEQ ID NO:9, GenBank Acc. No. M18405). Asterisks indicate amino acid residues that are conserved across all six species. This figure was created using the Pileup program of GCG (Program  
10 Manual for the Wisconsin Package, Version 9.0-OpenVMS, December 1996, Genetics Computer Group, 575 Science Drive, Madison, WI, USA 53711).

Figure 4 is a diagram describing the construction of the intermediate plasmid vector pT7BlueR + PDO1.

Figure 5 is a diagram describing the construction of *E. coli* expression vector pE24CP1.

15 Applicants have provided a sequence listing in conformity with "Rules for the Standard Representation of Nucleotide and Amino Acid Sequences in Patent Applications" (Annexes I and II to the Decision of the President of the EPO, published in Supplement No. 2 to OJ EPO, 12/1992) and with 37 C.F.R. 1.821-1.825 and Appendices A and B ("Requirements for Application Disclosures  
20 Containing Nucleotides and/or Amino Acid Sequences").

SEQ ID NO:1 presents a partial nucleic acid sequence of an expressed sequence tag (EST) bearing GenBank Accession No. T92052 obtained from an *Arabidopsis thaliana* cDNA library. This sequence was contained in clone 91B13T7 of the library.

25 SEQ ID NO:2 presents the initial determination of the nucleic acid sequence and the deduced amino acid sequence of a cDNA encoding a full-length form of *Arabidopsis thaliana p*-hydroxyphenylpyruvate dioxygenase enzyme, as contained in plasmid pGBPPD2.

SEQ ID NO:3 presents the initially deduced amino acid sequence encoded  
30 by a cDNA for *Arabidopsis thaliana p*-hydroxyphenylpyruvate dioxygenase enzyme.

SEQ ID NOS:4 and 5 present the nucleotide sequences of a pair of complementary oligonucleotides (CAM 32 and CAM 33, respectively) used to facilitate subcloning and expression of the gene encoding *p*-hydroxyphenyl-  
35 pyruvate dioxygenase without the chloroplast transit sequence.

SEQ ID NO:6 presents the amino acid sequence of *p*-hydroxyphenylpyruvate dioxygenase enzyme derived from human (GenBank Acc. No. U29895).

SEQ ID NO:7 presents the amino acid sequence of *p*-hydroxyphenyl-pyruvate dioxygenase enzyme derived from pig (GenBank Acc. No. D13390).

SEQ ID NO:8 presents the amino acid sequence of *p*-hydroxyphenyl-pyruvate dioxygenase enzyme derived from mouse (GenBank Acc. No. D29987).

5 SEQ ID NO:9 presents the amino acid sequence of *p*-hydroxyphenyl-pyruvate dioxygenase enzyme derived from rat (GenBank Acc. No. M18405).

SEQ ID NO:10 presents the nucleic acid sequence and deduced amino acid sequence of the cloned cDNA encoding the *Zea mays p*-hydroxyphenylpyruvate dioxygenase enzyme, as contained in plasmid pMPDO.

10 SEQ ID NO:11 presents the deduced amino acid sequence of the cloned cDNA encoding the *Zea mays p*-hydroxyphenylpyruvate dioxygenase enzyme, as contained in plasmid pMPDO.

SEQ ID NO:12 presents the nucleic acid sequence and the deduced amino acid sequence of the truncated form of *Arabidopsis thaliana p*-hydroxyphenyl-pyruvate dioxygenase enzyme as contained in pE24CP1.

15 SEQ ID NO:13 presents the deduced amino acid sequence of the truncated form of *Arabidopsis thaliana p*-hydroxyphenylpyruvate dioxygenase enzyme as contained in pE24CP1.

SEQ ID NO:14 presents the revised nucleic acid sequence and the deduced amino acid sequence of the cloned cDNA encoding the full-length *Arabidopsis thaliana p*-hydroxyphenylpyruvate dioxygenase enzyme, as contained in plasmid pGBPPD2.

20 SEQ ID NO:15 presents the revised amino acid sequence deduced from the cDNA for the full length *Arabidopsis thaliana p*-hydroxyphenylpyruvate dioxygenase enzyme.

25 SEQ ID NO:16 presents the nucleic acid sequence determined from a portion of a cDNA from *Vernonia galamensis*, as contained in clone vs1.pk0015.b2.

#### DETAILS OF THE INVENTION

##### BIOLOGICAL DEPOSITS

30 The following biological materials have been deposited under the terms of the Budapest Treaty at American Type Culture Collection (ATCC), 12301 Parklawn Drive, Rockville, MD 20852, and bear the following accession numbers:

35

<u>Depositor Identification</u>		<u>Int'l. Depository</u>	
<u>Host Strain</u>	<u>Plasmid</u>	<u>Accession Number</u>	<u>Date of Deposit</u>
<i>E. coli</i> BL21(DE3)	pE24CP1	ATCC 98083	June 25, 1996
N/A	pGBPPD2	ATCC 97622	June 25, 1996
N/A	pMPDO	ATCC 209120	June 12, 1997

### Definitions

In the context of this disclosure, a number of terms shall be utilized. As used herein, the term "nucleic acid" refers to a large molecule which can be single-stranded or double-stranded, composed of monomers (nucleotides) containing a sugar, phosphate and either a purine or pyrimidine. A "nucleic acid fragment" is a portion of a given nucleic acid molecule. As used herein, "DNA" (deoxyribonucleic acid) is the genetic material, whereas "RNA" (ribonucleic acid) is involved in the transfer of the information encoded by the DNA into proteins and polypeptides. A "genome" is the entire body of genetic material contained in each cell of an organism. The term "nucleotide sequence" refers to a polymer of DNA or RNA which can be single- or double-stranded, optionally containing synthetic, non-natural or altered nucleotide bases capable of incorporation into DNA or RNA polymers.

As used herein, "essentially similar" refers to DNA sequences that may involve base changes that do not cause a change in the encoded amino acid or which involve base changes which may alter one or more amino acids, but do not affect the functional properties of the protein encoded by the DNA sequence. It is therefore understood that the invention encompasses more than the specific exemplary sequences. Modifications to the sequence, such as deletions, insertions, or substitutions in the sequence which produce "silent changes" (i.e., those that do not substantially affect the functional properties of the resulting protein molecule) are also contemplated. For example, alteration(s) in the gene sequence which reflects the degeneracy of the genetic code, or which result in the production of a chemically equivalent amino acid at a given site, are contemplated; thus, a codon for the amino acid alanine, a hydrophobic amino acid, may be substituted by a codon encoding another less hydrophobic residue, such as glycine, or a more hydrophobic residue, such as valine, leucine, or isoleucine. Similarly, changes which result in substitution of one negatively charged residue for another, such as aspartic acid for glutamic acid, or one positively charged residue for another, such as lysine for arginine, can also be expected to produce a biologically equivalent product. Nucleotide changes which result in alteration of the N-terminal and C-terminal portions of the protein molecule would also not be



expected to alter the activity of the protein. In some cases, it may in fact be desirable to make mutants of the sequence in order to study the effect of alteration on the biological activity of the protein. Each of the proposed modifications is well within the routine skill in the art, as is determination of retention of  
5 biological activity of the encoded products. Moreover, the skilled artisan recognizes that "essentially similar" sequences encompassed by this invention are also defined by their ability to hybridize, under stringent conditions (0.1X SSC, 0.1% SDS, 65°C), with the sequences exemplified herein.

"Gene" refers to a nucleic acid fragment that encodes a specific protein,  
10 including regulatory sequences preceding (5' non-coding) and following (3' non-coding) the coding region. "Native" gene refers to the gene as found in nature with its own regulatory sequences. "Chimeric" gene refers to a gene comprising heterogeneous regulatory and coding sequences. "Endogenous" gene refers to the native gene normally found in its natural location in the genome. A "foreign"  
15 gene refers to a gene not normally found in the host organism but that is introduced by gene transfer.

"Coding sequence" refers to a DNA sequence that codes for a specific protein and excludes the non-coding sequences.

"Initiation codon" and "termination codon" refer to a unit of three adjacent  
20 nucleotides in a coding sequence that specifies initiation and termination, respectively, of protein synthesis (mRNA translation). "Open reading frame" refers to the amino acid sequence encoded between translation initiation and termination codons of a coding sequence.

"RNA transcript" refers to the product resulting from RNA polymerase-  
25 catalyzed transcription of a DNA sequence. When the RNA transcript is a perfect complementary copy of the DNA sequence, it is referred to as the primary transcript or it may be a RNA sequence derived from posttranscriptional processing of the primary transcript. "Messenger RNA" (mRNA) refers to RNA that can be translated into protein by the cell. "cDNA" refers to a double-stranded  
30 DNA, one strand of which is complementary to and derived from mRNA by reverse transcription. "Sense RNA" refers to RNA transcript that includes the mRNA.

As used herein, "regulatory sequences" are nucleotide sequences that control the transcription or expression of a coding sequence located upstream (5'), within,  
35 or downstream (3') to the coding sequence, act in conjunction with the protein biosynthetic apparatus of the cell and include promoters, translation leader sequences, transcription termination sequences, and polyadenylation sequences.

"Promoter" refers to a DNA sequence in a gene, usually upstream (5') to its coding sequence, which controls the expression of the coding sequence by providing the recognition for RNA polymerase and other factors required for proper transcription. A promoter may also contain DNA sequences that are involved in the binding of protein factors which control the effectiveness of transcription initiation in response to physiological or developmental conditions. In the case of eukaryotic organisms, it may also contain enhancer elements.

An "enhancer element" is a DNA sequence which can stimulate promoter activity. It may be an innate element of the promoter or a heterologous element inserted to enhance the activity level and tissue-specificity of a promoter. "Constitutive promoters" refer to those enhancer elements that direct gene expression in all tissues and at all times. "Organ-specific" or "development-specific" promoters as referred to herein are those that direct gene expression almost exclusively in specific organs, such as leaves or seeds, or at specific development stages in an organ, such as in early or late embryogenesis, respectively.

The term "operably linked" refers to nucleic acid sequences on a single nucleic acid molecule which are associated so that the function of one is affected by the other. For example, a promoter is operably linked with a structural gene (i.e., a gene encoding *p*-hydroxyphenylpyruvate dioxygenase, as disclosed herein) when it is capable of affecting the expression of that structural gene (i.e., that the structural gene is under the transcriptional control of the promoter).

The term "expression", as used herein, is intended to mean the production of the protein product encoded by a gene. More particularly, "expression" refers to the transcription and stable accumulation of the sense RNA (mRNA) derived from the nucleic acid fragment(s) of the invention that, in conjunction with the protein apparatus of the cell, results in altered levels of protein product.

"Overexpression" refers to the production of a gene product in transgenic organisms that exceeds levels of production in normal or non-transformed organisms. "Altered levels" refers to the production of gene product(s) in transgenic organisms in amounts or proportions that differ from that of normal or non-transformed organisms. "Facilitating expression" refers to steps and conditions for culturing host cells containing the desirable gene to yield an increased production of the enzyme. For example, addition of a chemical inducer specific to the particular promoter operably linked to the gene facilitates expression of the encoded enzyme. This is measured relative to the production levels of an untreated gene.

The "3' non-coding sequences" refers to the DNA sequence portion of a gene that contains a polyadenylation signal and any other regulatory signal capable of affecting mRNA processing or gene expression. The polyadenylation signal is usually characterized by affecting the addition of polyadenylic acid tracts to the 3' end of the mRNA precursor.

The "translation leader sequence" refers to that DNA sequence portion of a gene between the promoter and coding sequence that is transcribed into RNA and is present in the fully processed mRNA upstream (5') of the translation start codon. The translation leader sequence may affect processing of the primary transcript to mRNA, mRNA stability, or translation efficiency.

"Transformation" herein refers to the transfer of a foreign gene into the genome of a host organism and its genetically stable inheritance. Bacterial transformation can proceed by any of several methods well known in the art, including calcium chloride-mediated transformation and electroporation. Examples of methods of plant transformation include *Agrobacterium*-mediated transformation and particle-accelerated or "gene gun" transformation technology (U.S. Patent No. 4,945,050).

"Host cell" refers to the cell that is transformed with the introduced genetic material.

"Plasmid vector" refers to a double-stranded, closed circular, extra-chromosomal DNA molecule.

"Tolerant" or "tolerance" refers to a condition whereby a cell or an organism is able to withstand the effect of application of a compound or composition at a concentration or application rate that causes a demonstrable effect in or against cells or organisms that are not tolerant. For example, the growth or survival of a plant that is tolerant to application of a herbicidal compound or composition will be less affected than the growth or survival of a plant that is not tolerant to application of the herbicidal compound or composition.

#### Cloning of Plant Genes Encoding *p*-Hydroxyphenylpyruvate Dioxygenase

The *p*-hydroxyphenylpyruvate dioxygenases from plants are a promising new class of targets for new herbicidal compounds. In order to be able to study this enzyme in detail, and to have available supplies of enzyme for inhibitor screening, cDNA clones encoding plant *p*-hydroxyphenylpyruvate dioxygenases were identified. These nucleic acid fragments are useful for the production of their encoded enzymes, for isolation of clones from additional plant sources that encode other *p*-hydroxyphenylpyruvate dioxygenase enzymes, and for understanding the biochemical and structural properties of these enzymes.

Nucleic acid fragments comprising nucleotide sequences that encode different forms of the enzyme *p*-hydroxyphenylpyruvate dioxygenase from the plant *Arabidopsis thaliana* have now been isolated. Subsequently, these nucleotide sequences were expressed in *E. coli* cells and shown to direct the synthesis of plant *p*-hydroxyphenylpyruvate dioxygenase enzymes.

An automated search of nucleotide sequences contained in a database representing an *Arabidopsis* cDNA library for sequences homologous to other known, non-plant *p*-hydroxyphenylpyruvate dioxygenase genes revealed the plasmid cDNA clone 91B13T7. This cDNA was obtained from the Arabidopsis Seed Stock Center at Ohio State University. Plasmid DNA suitable for nucleotide sequence determination was prepared and the nucleotide sequence of the plasmid insert was determined. The resulting sequence was not interpretable, suggesting possible contamination of the plasmid sample by an extraneous nucleic acid. This assumption was confirmed by digesting the plasmid DNA sample with restriction enzymes and separating the resulting nucleic acid fragments by agarose gel electrophoresis. This analysis revealed the presence of nucleic acid fragments that could not be derived from the plasmid carrying the putative *p*-hydroxyphenylpyruvate dioxygenase fragment. Furthermore, a search of the publically available nucleic acid sequence databases revealed that the *Arabidopsis thaliana* sequence reported for cDNA clone 91B13T7 corresponded to a truncated cDNA (Figure 1). Based on publically available mammalian cDNA sequence information for *p*-hydroxyphenylpyruvate dioxygenase, the minimum length expected for a cDNA encoding a complete *p*-hydroxyphenylpyruvate dioxygenase enzyme is 1 kb (Table 1).

Table 1  
Predicted cDNA Length for Sequences  
Encoding *p*-Hydroxyphenylpyruvate Dioxygenase

Organism	Amino Acid Residues	Minimum cDNA (kb)
Human	392	1.176
Pig	392	1.176
<i>Pseudomonas</i> sp.	357	1.071

Therefore, based on the expected length of a cDNA capable of encoding a functional *p*-hydroxyphenylpyruvate dioxygenase, the *Arabidopsis thaliana* sequence obtained from the public database was insufficient to encode a full-length, active *p*-hydroxyphenylpyruvate dioxygenase enzyme. Therefore, a cDNA with the capacity to encode a full-length enzyme *Arabidopsis thaliana* was cloned,

as described herein. A 400 bp segment of the insert of plasmid 91B13T7 was liberated by digestion with restriction enzymes and used to screen a cDNA library prepared from norflurazon-treated *Arabidopsis thaliana* seedlings (Scolnik, P. A., and Bartley, G. E. (1994) *Plant Physiol.* 104:1469-1470). Several clones showing  
5 positive hybridization to this probe were sequenced. The initial determination of the sequence of the longest cDNA clone obtained from this effort is shown in Figure 2 and in SEQ ID NO:2. During the course of subsequent work with this clone it became necessary to confirm certain features of the sequence. A corrected sequence of this cDNA is presented in SEQ ID NO:12.

- 10 The sequence reported in Figure 2 indicates that this cDNA has the capacity to encode a protein of MW 48,841 which, as shown in Figure 3, has a high level of homology to *p*-hydroxyphenylpyruvate dioxygenase enzymes from other eukaryotes.

- 15 A cDNA capable of encoding a full-length *p*-hydroxyphenylpyruvate dioxygenase has also been obtained from corn. This cDNA, contained in plasmid pMPDO, was identified in a corn cDNA library using an approximately 900 base pairs portion of the *Arabidopsis* cDNA as a probe. The predicted amino acid sequence that is encoded by the corn cDNA is also compared to *p*-hydroxyphenylpyruvate dioxygenase enzymes from other eukaryotes in Figure 3.

- 20 A cDNA library was prepared from messenger RNA isolated from developing seeds of *Vernonia galamensis*. Random sequencing of the clones contained in the library identified a probable clone, designated vs1.pk0015.b2, for the *p*-hydroxyphenylpyruvate dioxygenase from this plant. The 513 bp expressed sequence tag (EST) is presented in SEQ ID NO:16.

- 25 Expression of the *Arabidopsis thaliana* cDNA Encoding *p*-Hydroxyphenylpyruvate Dioxygenase in *E. coli*

- The nucleic acid fragments of the instant invention encoding a plant *p*-hydroxyphenylpyruvate dioxygenase enzymes can be operably linked to suitable regulatory sequences, thereby creating chimeric genes that can be used to direct  
30 expression of the enzyme in transgenic organisms. These transgenic organisms include, but are not limited to: plants (*Plant Molecular Biology*; Croy, R. R. D., Ed.; Bios Scientific Publishers; 1993); microorganisms, including *Escherichia coli* (Gold, L. (1990) *Methods in Enzymology* 185:11), *Bacillus subtilis* (Henner, D. J. (1990) *Methods in Enzymology* 185:199), yeast (Gellissen, G., et al. (1992) *Antonie Leeuwenhoek* 62:79), and fungi, including members of the genus  
35 *Aspergillus* (Devchand, M. and Gwynne, D. I. (1991) *J. Biotechnol.* 17:3); and insect cells containing recombinant baculoviruses (Lukow, V. A. and Summers, M. D. (1988) *Bio/Technology* 6:47).

One skilled in the art can isolate the coding sequences from the fragments of the invention by using or creating sites for restriction endonucleases, as described in Sambrook, J., et al. ((1989) *Molecular Cloning, A Laboratory Manual*, 2nd ed.; Cold Spring Harbor Laboratory Press; hereinafter "Maniatis"). Alternatively, polymerase chain reaction (PCR) techniques can be employed to isolate and/or modify the fragments of the invention (Newton, C. R. and Graham, A. (1994) *PCR*; Bios Scientific Publishers).

*Arabidopsis p*-hydroxyphenylpyruvate dioxygenase was expressed in *E. coli* under control of a T7 promoter in a strain expressing T7 RNA polymerase (Studier, F. W., et al. (1990) *Methods in Enzymology* 185:60). Promoters other than T7 are commonly used in expression vectors and could be substituted for protein expression in *E. coli*. Examples of alternative promoters include, but are not limited to, *trp* (Yansura, D. G. and Henner, D. J. (1990) *Methods in Enzymology* 185:54),  $P_L$  (Remaut, E. et al. (1981) *Gene* 15:81), *tac* (Amann, E. et al. (1983) *Gene* 25:167), *trc* (Amann, E. et al. (1988) *Gene* 69:301), and promoters such as *lacUV5*, *lpp*,  $P_R$ , and hybrid and tandem promoters constructed to combine specific features to increase strength or regulation capacity (Balbas, P. and Bolivar, F. (1990) *Methods in Enzymology* 185:14).

#### Biochemical Evidence of Enzymatic Function

The enzyme *p*-hydroxyphenylpyruvate dioxygenase catalyzes the reaction of *p*-hydroxyphenylpyruvate with molecular oxygen to give homogentisate and CO<sub>2</sub>. The enzyme can be assayed by measuring oxygen utilization (Hager, S. E., et al. (1957) *J. Biol. Chem.* 225:935-947), CO<sub>2</sub> release or homogentisate production from radioactive labeled *p*-hydroxyphenylpyruvate (Lindblad, B. (1971) *Clin. Chem. Acta* 34:113-121), loss of the *p*-hydroxyphenylpyruvate (Lin, E. C. C. et al. (1958) *J. Biol. Chem.* 233:668-673), or formation of homogentisate using a colorimetric assay (Fellman, J. H. et al. (1972) *Biochim. Biophys. Acta* 284:90-100) or UV detection following HPLC or a similar chromatographic separation technique. The activity of *p*-hydroxyphenylpyruvate dioxygenase may also be measured in a coupled assay in which the initial product, homogentisate, is oxidized by homogentisate dioxygenase; formation of maleylacetoacetate determined by measuring absorbance at 330 nm (Fernández-Cañón, J. M. and Peñalva, M. A. (1997) *Anal. Biochem.* 245:218-221).

An alternative to any of the kinetic assays for *p*-hydroxyphenylpyruvate dioxygenase is an end-point or fixed-time assay. The procedure is based on the conversion of unconverted substrate, *p*-hydroxyphenylpyruvate to its enediol tautomer by tautomerase in the presence of borate ions and measurement of the characteristic 308 nm peak of the tautomer (Lin, E. C. C. et al. (1958) *J. Biol.*

*Chem.* 233:668-673). The procedure involves the addition of enough *p*-hydroxyphenylpyruvate dioxygenase to consume ~80% of the organic substrate over 1 hour in 200  $\mu$ L of assay buffer, which in this case is a 50 mM Tris, pH 7.4, 0.10 mM *p*-hydroxyphenylpyruvic acid, 1.75 mM ascorbate and 1.25 mM EDTA.

5 After 1 hr the reaction is quenched by the addition of 100  $\mu$ L of 0.8 M borate, pH 7.3, containing 1000 ppb of a *p*-hydroxyphenylpyruvate dioxygenase inhibitor and 0.25  $\mu$ L of 6.1 mg/mL of tautomerase. The absorbance at 308 nm is read after a 30 min incubation and is stable thereafter for 2 hr. The advantage of this assay over the kinetic procedure is that the *p*-hydroxyphenylpyruvate dioxygenase is not  
10 required to oxidize the substrate in the presence of high concentrations of borate, a condition that might interfere with the mode of action of inhibitors. Furthermore the assay produces essentially a stable binary indication of *p*-hydroxyphenylpyruvate dioxygenase inhibition, and is well-suited for applications which require a high-throughput of samples and assays.

15 The enzyme encoded by the nucleic acid fragments and overexpressed in *E. coli* can be extracted in any conventional buffer used for extracting soluble plant enzymes. Although a large amount of an overexpressed protein is often insoluble, the amount that is soluble represents can represent as much as 50% of the total soluble protein. Soluble overexpressed protein has high *p*-hydroxyphenylpyruvate dioxygenase activity and is easily extracted. Likewise, it may be  
20 possible to resolubilize an insoluble overexpressed protein in an active form under appropriate conditions, since addition of sarkosyl (sodium N-lauroylsarcosinate) to the extraction buffer appeared to increase the amount of the overexpressed protein extracted. For optimum activity, a reducing agent such as ascorbate or  
25 reduced glutathione should be present as well as a source a ferrous ion.

An overexpressed enzyme can be assayed using all the techniques described above for measuring *p*-hydroxyphenylpyruvate dioxygenase activity, while only the techniques using labeled *p*-hydroxyphenylpyruvate can be used to measure activity in crude plant extracts. Therefore, the availability of an  
30 overexpressed enzyme greatly facilitates the development of high capacity screens to identify inhibitors of the enzyme. Potential inhibitors are evaluated for their capacity to reduce the rate of the reaction of the enzyme, resulting in reduced oxygen uptake and CO<sub>2</sub> release, and lower rates of formation of homogentisate and loss of *p*-hydroxyphenylpyruvate. Applicants have demonstrated that at least  
35 one of the instant nucleic acid fragments can be overexpressed in *E. coli* cells, resulting in production of a protein that catalyzes the conversion of *p*-hydroxyphenylpyruvate to homogentisate with the release of CO<sub>2</sub>. Furthermore, it has been shown that this activity is inhibited by commercial herbicides known to

inhibit *p*-hydroxyphenylpyruvate dioxygenase. Finally, an overexpressed enzyme can be used in a high capacity assay to identify compounds that inhibit the enzymatic activity of *p*-hydroxyphenylpyruvate dioxygenase. Such compounds may serve as herbicides.

5 Preparation of Plants Tolerant to Inhibitors of *p*-Hydroxyphenylpyruvate Dioxygenase

This invention embodies plants which are resistant or at least tolerant to herbicides that target the *p*-hydroxyphenylpyruvate dioxygenase enzyme at levels which are normally inhibitory to the naturally occurring *p*-hydroxyphenylpyruvate  
10 dioxygenase enzyme. This altered *p*-hydroxyphenylpyruvate dioxygenase activity is conferred by (1) overexpression of the wild-type *p*-hydroxyphenylpyruvate dioxygenase enzyme, or (2) expression of a DNA molecule encoding a herbicide-tolerant enzyme. The said enzyme may be a modified form of an *p*-hydroxy-phenylpyruvate dioxygenase enzyme that occurs naturally in a eukaryote or  
15 prokaryote, or a modified form of an *p*-hydroxyphenylpyruvate dioxygenase enzyme that naturally occurs in a plant, or a herbicide tolerant enzyme that naturally occurs in a prokaryote (Duke et al. *Herbicide Resistant Crops*; Lewis: Boca Raton;1994). An effective amount of gene expression to render the cells of the plant tissue substantially tolerant to the herbicide depends on whether the gene  
20 codes for an unaltered *p*-hydroxyphenylpyruvate dioxygenase gene or a mutant or altered form of the gene that is less sensitive to the herbicides. Expression of an unaltered plant *p*-hydroxyphenylpyruvate dioxygenase gene in an effective amount is that amount that provides for a 2- to 10-fold increase in herbicide tolerance. Plants encompassed by the invention include monocotyledoneous and  
25 dicotyledoneous plants. Preferred are those plants which would be potential targets for *p*-hydroxyphenylpyruvate dioxygenase-inhibiting herbicides, particularly agronomically important crops such as maize and other cereal crops.

Increased levels of expression of *p*-hydroxyphenylpyruvate dioxygenase activity, from two to ten or more times the natively expressed amount, would be  
30 sufficient to overcome growth inhibition caused by the herbicide. Plants containing such altered *p*-hydroxyphenylpyruvate dioxygenase enzyme activity can be obtained by direct selection in plants. This method is known in the art. See, e.g., U.S. Patent No. 5,162,602, U.S. Patent No. 4,761,373, and references cited therein.

35 Overexpression of *p*-hydroxyphenylpyruvate dioxygenase also can be accomplished by stably transforming a host plant cell with a chimeric DNA molecule comprising a promoter capable of driving expression of an associated coding sequence in a plant cell and operably linked to a homologous or



heterologous coding sequence encoding *p*-hydroxyphenylpyruvate dioxygenase.

A "homologous" *p*-hydroxyphenylpyruvate dioxygenase gene is isolated from an organism taxonomically identical to the target plant cell, whereas a "heterologous" *p*-hydroxyphenylpyruvate dioxygenase gene is obtained from an organism taxonomically distinct from the target plant.

The expression of foreign genes in plants is well-established (De Blaere et al., (1987) *Meth. Enzymol.* 143:277-291). Promoters utilized to drive gene expression in transgenic plants or plant cells (i.e., those capable of driving expression of the associated coding sequences such as *p*-hydroxyphenylpyruvate dioxygenase in plant cells, include those directing the 19S and 35S transcripts in Cauliflower mosaic virus (Odell et al., (1985) *Nature* 313:810-812; Hull et al., (1987) *Virology* 86:482-493), small subunit of ribulose 1,5-bisphosphate carboxylase (Morelli et al., (1985) *Nature* 315:200-204; Broglie et al., (1984) *Science* 224:838-843; Hererra-Estrella et al., (1984) *Nature* 310:115-120; Coruzzi et al., (1984) *EMBO J.* 3:1671-1679; Faciotti et al., (1985) *Bio/Technology* 3:241 and chlorophyll a/b binding protein (Lamppa et al., (1986) *Nature* 316:750-752); nopaline synthase promoters (Depicker et al. (1982) *J. Mol. App. Genet.* 1:561-573; An et al. (1990) *Plant Cell* 2:225-233). The chimeric DNA construct(s) of the invention may contain multiple copies of a promoter or multiple copies of the *p*-hydroxyphenylpyruvate dioxygenase coding sequences. In addition, the construct(s) may include coding sequences for selectable markers and coding sequences for other peptides such as signal or transit peptides. The preparation of such constructs is within the ordinary level of skill in the art. Resistance to inhibitors of the plant carotenoid biosynthesis pathway, which is also targeted by *p*-hydroxyphenylpyruvate dioxygenase inhibitors, has been achieved by expressing a bacterial gene encoding phytoene desaturase driven by the CaMV promoter (Misawa et al., (1994) *Plant. J.* 4:481-490).

Transit peptides may be fused to the *p*-hydroxyphenylpyruvate dioxygenase coding sequence in the chimeric DNA constructs of the invention to direct transport of the expressed *p*-hydroxyphenylpyruvate dioxygenase enzyme to the desired site of action. Examples of transit peptides include the chloroplast transit peptides such as those described in Von Heijne et al., (1991) *Plant Mol. Biol. Rep.* 9:104-126; Mazur et al., (1987) *Plant Physiol.* 85:1110; Vorst et al., (1988) *Gene* 65:59; and mitochondrial transit peptides such as those described in Boutry et al., (1987) *Nature* 328:340-342.

It is envisioned that the introduction of enhancers or enhancer-like elements into other promoter constructs will also provide increased levels of primary transcription to accomplish the invention. These would include viral enhancers

such as that found in the 35S promoter (Odell et al., (1988) *Plant Mol. Biol.* 10:263-272), enhancers from the opine genes (Fromm et al., (1989) *Plant Cell* 1:977-984), or enhancers from any other source that result in increased transcription when placed into a promoter operably linked to the nucleic acid  
5 fragment of the invention.

Introns isolated from the maize Adh-1 and Bz-1 genes (Callis et al., (1987) *Genes Dev.* 1:1183-1200), and intron 1 and exon 1 of the maize Shrunken-1 (sh-1) gene (Maas et al., (1991) *Plant Mol. Biol.* 16:199-207) may also be of use to increase expression of introduced genes. Results with the first intron of the maize  
10 alcohol dehydrogenase (Adh-1) gene indicate that when this DNA element is placed within the transcriptional unit of a heterologous gene, mRNA levels can be increased by 6.7-fold over normal levels. Similar levels of intron enhancement have been observed using intron 3 of a maize actin gene (Luehrsen, K. R. and Walbot, V., (1991) *Mol. Gen. Genet.* 225:81-93). Enhancement of gene  
15 expression by Adh1 intron 6 (Oard et al., (1989) *Plant Cell Rep* 8:156-160) has also been noted. Exon 1 and intron 1 of the maize sh-1 gene have been shown to individually increase expression of reporter genes in maize suspension cultures by 10 and 100-fold, respectively. When used in combination, these elements have been shown to produce up to 1000-fold stimulation of reporter gene expression  
20 (Maas et al., (1991) *Plant Mol. Biol.* 16:199-207).

Any 3' non-coding region capable of providing a polyadenylation signal and other regulatory sequences that may be required for proper expression can be used to accomplish the invention. This would include the 3' end from any storage protein such as the 3' end of the 10kd, 15kd, 27kd and alpha zein genes, the 3' end  
25 of the bean phaseolin gene, the 3' end of the soybean  $\beta$ -conglycinin gene, the 3' end from viral genes such as the 3' end of the 35S or the 19S cauliflower mosaic virus transcripts, the 3' end from the opine synthesis genes, the 3' ends of ribulose 1,5-bisphosphate carboxylase or chlorophyll a/b binding protein, or 3' end sequences from any source such that the sequence employed provides the  
30 necessary regulatory information within its nucleic acid sequence to result in the proper expression of the promoter/coding region combination to which it is operably linked. There are numerous examples in the art that teach the usefulness of different 3' non-coding regions (for example, see Ingelbrecht et al., (1989) *Plant Cell* 1:671-680).

35 Various methods of introducing a DNA sequence (i.e., of transforming) into eukaryotic cells of higher plants are available to those skilled in the art (see EPO publications 0 295 959 A2 and 0 138 341 A1). Such methods include high-velocity ballistic bombardment with metal particles coated with the nucleic acid

constructs (see Klein et al., (1987) *Nature* (London) 327:70-73. and see U.S. Patent No. 4,945,050), as well as those based on transformation vectors based on the Ti and Ri plasmids of *Agrobacterium* spp., particularly the binary type of these vectors. Ti-derived vectors transform a wide variety of higher plants, including

5 monocotyledonous and dicotyledonous plants, such as soybean, cotton and rape seed (Pacciotti et al., (1985) *Bio/Technology* 3:241; Byrne et al., (1987) *Plant Cell, Tissue and Organ Culture* 8:3; Sukhapinda et al., (1987) *Plant Mol. Biol.* 8:209-216; Lorz et al., (1985) *Mol. Gen. Genet.* 199:178-182; Potrykus et al., (1985) *Mol. Gen. Genet.* 199:183-188).

10 Other transformation methods are available to those skilled in the art, such as direct uptake of foreign DNA constructs (see EPO publication 0 295 959 A2), and techniques of electroporation (see Fromm et al., (1986) *Nature* (London) 319:791-793). Once transformed, the cells can be regenerated by those skilled in the art. Also relevant are several recently described methods of introducing

15 nucleic acid fragments into commercially important crops, such as rapeseed (see De Block et al., (1989) *Plant Physiol.* 91:694-701), sunflower (Everett et al., (1987) *Bio/Technology* 5:1201-1204), soybean (McCabe et al., (1988) *Bio/Technology* 6:923-926; Hinchey et al., (1988) *Bio/Technology* 6:915-922; Chee et al., (1989) *Plant Physiol.* 91:1212-1218; Christou et al., (1989) *Proc. Natl. Acad. Sci USA* 86:7500-7504; EPO Publication 0 301 749 A2), and corn (Gordon-Kamm et al., (1990) *Plant Cell* 2:603-618; and Fromm et al., (1990) *Bio/Technology* 8:833-839).

Altered *p*-hydroxyphenylpyruvate dioxygenase enzyme activity may also be achieved through the generation or identification of modified forms of the isolated

25 eukaryotic *p*-hydroxyphenylpyruvate dioxygenase coding sequence having at least one amino acid substitution, addition or deletion which encodes an altered *p*-hydroxyphenylpyruvate dioxygenase enzyme resistant to a herbicide that inhibits the unaltered, naturally occurring form. Genes encoding such enzymes can be obtained by numerous strategies known in the art. A first general strategy

30 involves direct or indirect mutagenesis procedures on microbes (e.g., *E. coli*, *S. cerevisiae* (Miller, (1972) *Experiments in Molecular Genetics*, Cold Spring Harbor Laboratory, Cold Spring Harbor, NY; Davis et al., (1980) *Advanced Bacterial Genetics*, Cold Spring Harbor Laboratory, Cold Spring Harbor, NY; Sherman et al., (1983) *Methods in Yeast Genetics*, Cold Spring Harbor Laboratory, Gold Spring Harbor NY; and U.S. Patent No. 4,975,374) and

35 cyanobacteria (Bryant, *The Molecular Biology of Cyanobacteria*; Kluwer Academic Publishers: Boston, 1995). A second method of obtaining mutant herbicide-resistant alleles of the eukaryotic *p*-hydroxyphenylpyruvate dioxygenase

enzyme involves direct selection in plants. For example, the effect of inhibitors on the growth of plants such as *Arabidopsis*, soybean, or maize may be determined by plating seeds sterilized by art-recognized methods on plates on a simple minimal salts medium containing increasing concentrations of the inhibitor. The lowest dose at which significant growth inhibition can be reproducibly detected is used for subsequent experiments. Mutagenesis of plant material may be utilized to increase the frequency at which resistant alleles occur in the selected population. Mutagenized seed material can be derived from a variety of sources, including chemical or physical mutagenesis of seeds, or chemical or physical mutagenesis or pollen (Neuffer, In *Maize for Biological Research*. Sheridan, ed. Univ. Press, Grand Forks, ND., pp. 61-64 (1982)), which is then used to fertilize plants and the resulting M1 mutant seeds collected. Typically, for *Arabidopsis*, M2 seeds (i.e., progeny seeds of plants grown from seeds mutagenized with chemicals, such as ethyl methane sulfonate, or with physical agents, such as gamma rays or fast neutrons) are plated at densities of up to 10,000 seeds/plate (10 cm diameter) on minimal salts medium containing an appropriate concentration of inhibitor. Seedlings that continue to grow and remain green 7-21 days after plating are transplanted to soil and grown to maturity and seed set. Progeny of these seeds are tested for resistance to the herbicide. If the resistance trait is dominant, plants whose seed segregate 3:1 (resistant:sensitive) are presumed to have been heterozygous for the resistance at the M2 generation. Plants that give rise to all resistant seed are presumed to have been homozygous for the resistance at the M2 generation. Such mutagenesis on intact seeds and screening of their M2 progeny seed can also be carried out on other species, for instance soybean (see, e.g., U.S. Patent No. 5,084,082). Mutant seeds to be screened for herbicide tolerance can also be obtained as a result of fertilization with pollen mutagenized by chemical or physical means.

#### EXAMPLE 1

##### Cloning of a cDNA for *Arabidopsis thaliana*

##### *p*-Hydroxyphenylpyruvate Dioxygenase

The plasmid containing the *Arabidopsis thaliana* 91B13T7 expressed sequence tag (Newman et al., (1994) *Plant Physiol* 106:1241-1255) was digested with the restriction enzymes *Bam*HI and *Eco*RI, and the resulting 400 bp fragment was used to screen a lambda phage cDNA library of *Arabidopsis thaliana* seedlings (Scolnik, P. A. and Bartley, G. E. (1994) *Plant Physiol*. 104:1469-1470) according to the following protocol.

*E. coli* KW251 cells were grown overnight in Luria Broth ("LB") containing 0.2% maltose and 10 mM MgSO<sub>4</sub>. Cells were pelleted by centrifugation and

resuspended in 10 mM MgSO<sub>4</sub> to an OD<sub>600</sub> of 0.5. Cell aliquots (0.8 mL) were mixed with 0.1 mL of diluted phage samples and 7 mL of top agarose (0.7% agarose in LB containing 10 mM MgSO<sub>4</sub>) at 45°C, and plated onto 150 mm Petri dishes containing LB agar. Phage plaques became visible in 5-7 h, at which point the plates were placed at 4°C.

Phage plaques were transferred to nitrocellulose filters according to standard techniques, and the filters were hybridized to <sup>32</sup>P-radiolabeled probe prepared according to the method of Feinberg and Vogelstein ((1983) *Anal. Biochem.* 132:6-13), using the hybridization conditions of Berlyn et al.((1989) *Proc. Natl. Acad. Sci.* 86:4604-4608). After exposure to X-ray film for 48 h, 12 positive plaques were eluted, plated, and hybridized under the same conditions. A total of 9 plaques that retained positive signals in this second round of hybridization were subjected to *in vivo* excision using the Exassist/SOLR™ system according to the manufacturer's protocol (Stratagene Cloning Systems, La Jolla, CA). DNA from the plasmids resulting from *in vivo* excision of positive plaques was prepared for DNA sequencing using the Wizard Plus™ kit (Promega, Madison, WI). Eight of the clones that were sequenced showed strong conservation with available *p*-hydroxyphenylpyruvate dioxygenase sequences, whereas the remaining clone did not correspond to a *p*-hydroxyphenylpyruvate dioxygenase. Alignment with known *p*-hydroxyphenylpyruvate dioxygenase sequences also revealed that two of the clones correspond to 0.3 kbp fragments from the 3' end of the transcript, and another two to 1.2 kbp fragments from the 5' end of the transcript. One clone of each was used to assemble a 1.5 kbp cDNA by ligating at the internal *Nhe*I restriction site (Figure 1). The initial determination of the DNA sequence (SEQ ID NO:2) of the resulting cDNA clone is shown in Figure 2. Subsequent work with this DNA fragment required confirmation of some of the features of its sequence. Approximately ten nucleotide residues were found to have been listed in error. Thus a corrected sequence for this DNA fragment is listed in SEQ ID NO:14 and the deduced amino acid sequence is set forth in SEQ ID NO:15. The revised sequences form the bases for analyses and comparisons reported herein.

#### EXAMPLE 2

##### Overexpression of the *Arabidopsis* cDNA in *E. coli*

The deduced amino acid sequence for *Arabidopsis* *p*-hydroxyphenylpyruvate dioxygenase was aligned with the amino acid sequences of *p*-hydroxyphenylpyruvate dioxygenase from mouse, pig, and *Streptomyces avermitilis* using the Pileup program of GCG (Program Manual for the Wisconsin Package, Version 8, September 1994, Genetics Computer Group, 575 Science Drive, Madison, WI, USA 53711). This analysis suggested an additional

29 amino acid-extension at the amino terminus of the *Arabidopsis* sequence (positions 1-29, Figure 3 and SEQ ID NO:3). This amino-terminal extension was assumed to be a chloroplast transit peptide which would be absent from the mature enzyme. Therefore, removal of the chloroplast transit peptide coding sequence coincided with transfer of the *p*-hydroxyphenylpyruvate dioxygenase coding sequence from the cloning vector into the expression vector.

The *Arabidopsis p*-hydroxyphenylpyruvate dioxygenase cDNA was moved from the pBluescript SK- cloning vector (Stratagene, La Jolla, CA) to the pET24c(+) expression vector (Novagen, Madison, WI) through the intermediate cloning vector pT7BlueR (Novagen). The plasmid pGBPPD2 consists of the *Arabidopsis p*-hydroxyphenylpyruvate dioxygenase cDNA and the pBluescript SK- cloning vector (Stratagene). The plasmid pE24CP1 consists of the *Arabidopsis p*-hydroxyphenylpyruvate dioxygenase cDNA, without the putative chloroplast transit peptide DNA sequence, and the pET24c(+) expression vector (Novagen).

The plasmids pGBPPD2 and pT7BlueR (5 µg each) were individually digested with 20 units of Xba I (New England Biolabs, NEB, Beverly, MA) and 20 units of Hind III (Gibco BRL, Gaithersburg, MD) in NEB restriction enzyme buffer 2 supplemented with 100 µg/mL bovine serum albumin at 37 °C for 1.75 h. Digesting pGBPPD2 with the restriction enzymes Xba I and Hind III releases the 5' and 3' ends, respectively, of the *p*-hydroxyphenylpyruvate dioxygenase cDNA from the pBluescript SK- polylinker. Products of the digestion were electrophoretically separated in a 1 percent agarose gel using TRIS/acetate/EDTA (TAE) buffer and visualized with ethidium bromide staining (Maniatis). Digestion of pGBPPD2 with the two restriction endonucleases resulted in a 2922 bp vector band and 1499 bp *p*-hydroxyphenylpyruvate dioxygenase cDNA band. Only a 2863 bp band was apparent after digesting pT7BlueR with the two enzymes, although a 24 bp fragment would also result. The 1499 bp *p*-hydroxyphenylpyruvate dioxygenase band and the 2863 bp T7BlueR band were cut out of the gel and the associated DNA purified from the agarose using a QIAquick Gel Extraction Kit (Qiagen, Chatsworth, CA) according to the manufacturer's instructions. The purified DNA samples were precipitated by the addition of sodium acetate (pH 5.2) to 0.3 M, 10 µg tRNA (added as carrier), two volumes of -20 °C ethanol and incubation at -20 °C overnight. Nucleic acid pellets were collected by centrifugation, washed with 70% ethanol and air dried. Both pellets were solubilized in 10 µL of TRIS/EDTA (TE) buffer, pH 8 (Maniatis), and then 1 µL of each sample loaded onto a 1% agarose, TAE gel in separate wells next to a well containing 4 µL of Mass Ladder (Gibco BRL). All samples were adjusted

to 10  $\mu$ L with water before loading. DNA was quantified by comparing band intensities of each sample with Mass Ladder band intensities following ethidium bromide staining and UV illumination.

Approximately 300 ng of *p*-hydroxyphenylpyruvate dioxygenase insert was mixed with 300 ng of double digested pT7BlueR vector in a total volume of 7  $\mu$ L and then heated to 45 °C for 5 min followed by cooling on ice. T4 DNA ligase buffer (Gibco BRL) and 1 unit of T4 DNA ligase (Gibco BRL) were added to the cooled DNA for a total volume of 10  $\mu$ L. The ligation mix was incubated at room temperature for 4 h and then transformed into MAX Efficiency DH5 $\alpha$  Competent Cells (Gibco BRL) of *E. coli* according to standard procedures (Maniatis). Transformed bacteria were spread onto LB agar plates supplemented with 100  $\mu$ g/mL carbenicillin and incubated overnight at 37 °C. Seventeen bacterial colonies were selected for subsequent analysis. A portion of each colony was inoculated into a separate 17x100 mm polypropylene culture tube (Falcon, Lincoln Park, NJ) containing 2 mL of liquid LB media and 200  $\mu$ g/mL carbenicillin. Liquid bacteria cultures were incubated overnight at 37 °C with shaking (250 rpm). Plasmid DNA was then isolated using a QIAprep Spin Plasmid Miniprep Kit (Qiagen) according to the manufacturer's instructions. A portion (5  $\mu$ L out of 50  $\mu$ L total) of each plasmid preparation was digested with 10 units each of Hind III and EcoR V (Gibco BRL) in a total volume of 15  $\mu$ L with React 2 buffer (Gibco BRL) for one h. (Note: The EcoRV site in the pBluescript polylinker was destroyed during the preparation of pGBPPD2 so only the EcoRV site in the pT7BlueR polylinker would be accessible to the restriction nuclease). Samples were separated electrophoretically in 1% agarose and tris/borate/EDTA (TBE) buffer (Maniatis). Bands were visualized with ethidium bromide staining; 7 out of 17 samples which contained 2 bands (2837 and 1525 bp) contained the *p*-hydroxyphenylpyruvate dioxygenase insert and were designated pT7BlueR+PDO1 (see Figure 4).

In order to remove the putative chloroplast transit sequence, the remaining 45  $\mu$ L of each prep of pT7BlueR+PDO1 were combined into a single sample and the DNA content determined spectrophotometrically at A<sub>260</sub> (Maniatis). A portion (5  $\mu$ g) of pT7BlueR+PDO1 was digested with 16 units of Eco47 III (MBI Fermentas) in a total volume of 100  $\mu$ L containing buffer 0 (MBI Fermentas) at 37 °C for 2 h. The digested plasmid DNA was then precipitated with sodium acetate and ethanol as above and the resulting dried nucleic acid pellet was dissolved in 60  $\mu$ L of React 2 (Gibco BRL) containing 20 units of Nde I (Gibco BRL) and incubated 2 h at 37 °C. The double digested sample was then loaded onto a 1% agarose gel in TAE and the large 4166 bp Nde I-Eco47III fragment

separated from the 196 bp fragment electrophoretically. The large fragment was cut out of the gel, purified from agarose and precipitated as above.

An oligonucleotide mix was prepared consisting of 100 pmoles each of oligos CAM32 and CAM33 (SEQ ID NOS:4 and 5, respectively) in a combined volume of 9.9  $\mu$ L. The two oligos complement each other to form a 3' blunt end corresponding to the 5' half of an Eco47 III restriction site and also form a 5' staggered end which corresponds to the 3' half of an Nde I restriction site.

CAM 32: (SEQ ID NO:4)

10 5'-TATGTCCAAGTTCGTAAGAAAGAATCCAAAGTCTGATAAATTCAAGGTTAAGC-3'

CAM 33: (SEQ ID NO:5)

5'-GCTTAACCTTGAATTTATCAGACTTTGGATTCTTCTTACGAACTTGGACA-3'

15 The oligo mix was heated to 90 °C for 1.5 min and then allowed to cool to room temperature over 20 min. The dried nucleic acid pellet resulting from purification of the 4166 bp Nde I-Eco47 III fragment was solubilized in 7  $\mu$ L of the cooled oligo mix and subsequently heated to 45 °C for 5 min followed by cooling on ice. Ligation of the oligos with the Nde I-Eco47 III fragment followed  
20 by transformation into DH5 $\alpha$  was performed as above. Transformed bacterial cells were spread onto LB/carbenicillin plates and incubated at 37 °C overnight. Seventeen colonies were selected and processed to isolate plasmid DNA as above. A portion (5 out of 50  $\mu$ L) of each plasmid was double digested with 10 units each of Nde I and Hind III and the fragments separated electrophoretically on a 1%  
25 agarose gel in TBE. A two band pattern corresponding to insert (1373 or 1518 bp) and vector (2844 bp) was detected. An additional double digest with 10 units each of Xba I and Hind III was performed on another 5  $\mu$ L aliquot of plasmids. When digested with Nde I and Hind III, none of the plasmids which contained the smaller insert size contained a Xba I site. The Xba I site would be eliminated if  
30 the two oligos replaced the 196 bp fragment originally present in pT7Blue+PDO1. The 7 plasmid samples with the modified *p*-hydroxyphenylpyruvate dioxygenase insert were combined and designated pT7BlueR+PDO2.

The pT7BlueR+PDO2 plasmid DNA was quantified spectrophotometrically (above) and then 5  $\mu$ g was digested with 20 units each of Hind III and Nde I in  
35 62  $\mu$ L of React 2 for 2 h at 37 °C. The digested sample was subsequently loaded onto a 1% agarose gel in TAE and separated electrophoretically. The 1373 bp fragment was isolated and precipitated as above. The plasmid pET24c(+) (5  $\mu$ g) was double digested with 20 units each of both Nde I and Hind III in React 2 at 37 °C for 2 h and the 5245 bp fragment then gel purified on a 1% agarose gel in



TAE and subsequently separated from agarose and precipitated as above. The dried pET24c(+) pellet was solubilized in 10  $\mu$ L TE and then 8  $\mu$ L was adjusted to a 20  $\mu$ L total volume with water, dephosphorylation buffer (Gibco BRL) and 1 unit of calf intestinal alkaline phosphatase (Gibco BRL). The sample was

5 incubated at 37 °C for 30 min and then gel purified, separated from agarose, and precipitated as above. The dried, dephosphorylated, pET24c(+) vector pellet and modified *p*-hydroxyphenylpyruvate dioxygenase insert pellet were each solubilized in 10  $\mu$ L TE and then 1  $\mu$ L of each was run on a 1% agarose TBE gel with 4  $\mu$ L of mass ladder to quantify DNA as above. One hundred nanograms of modified

10 *p*-hydroxyphenylpyruvate dioxygenase insert was mixed with 120 ng of dephosphorylated pET24c(+) vector in a total of 7  $\mu$ L volume. The mix was heated to 45 °C for 5 min and then cooled on ice. The mix was then supplemented with T4 DNA ligase buffer and 1 unit of T4 DNA ligase in a total volume of 10  $\mu$ L and the mix allowed to incubate at room temperature for 4 h. The ligation

15 mix was subsequently transformed into DH5 $\alpha$ , spread on LB agar supplemented with 30  $\mu$ g/mL kanamycin, and incubated overnight at 37 °C. Plasmid preparations were performed on 11 colonies as above. Plasmids were double digested with Nde I and Hind III and fragments separated electrophoretically. All plasmids had the expected 1373 bp and 5245 bp fragments. One bacteria colony

20 was selected and used to inoculate 100 mL of liquid LB supplemented with 30  $\mu$ g/mL kanamycin which was subsequently incubated at 37 °C overnight with shaking. Plasmid DNA was isolated from the resulting bacteria culture using a Qiagen Plasmid Midi Kit according to the manufacturer's instructions. A portion of the plasmid DNA (pE24CP1) was sequenced with the Sequenase Version 2.0

25 DNA Sequencing Kit (United States Biochemical, Cleveland, OH) using a biotinylated sequencing primer to the T7 promoter (United State Biochemical) according to the manufacturer's instructions for non-radioactive manual sequencing. DNA was transferred from the sequencing gel to Hybond-N+ nylon transfer membrane (Amersham, Arlington Heights, IL) by capillary action.

30 Transfer and all subsequent steps in chemiluminescent detection of DNA fragments were performed with a SEQ-Light Chemiluminescent Sequencing System kit (Tropix, Bedford, MA) according to the manufacturer's instructions. DNA sequencing verified that the plasmid contained the expected 5' sequence for the modified *p*-hydroxyphenylpyruvate dioxygenase insert where nucleotides 1-95

35 (Figure 2) were replaced with an ATG transcriptional start site. This is equivalent to amino acids 2-29 (Figure 3) being eliminated from the N-terminus of the *Arabidopsis p*-hydroxyphenylpyruvate dioxygenase amino acid sequence.

The plasmid pE24CP1 was transformed into competent cells of BL21(DE3) *E. coli* (Novagen), as above. Transformed cells were spread on LB/kanamycin plates and incubated overnight at 37 °C. Seven colonies were selected for plasmid preparations as above and plasmid DNA was double digested with Nde I and Hind III to verify that all plasmids had the expected electrophoretic banding pattern. One colony was selected and streaked for isolation on LB/kanamycin plates. A well isolated colony was used to inoculate liquid LB supplemented with 30 µg/mL kanamycin and the culture was incubated at 37 °C with shaking (250 rpm) until it reached an A<sub>600</sub> of 0.6 absorbance units. An 8% glycerol freezer stock was prepared according to the Novagen protocol and stored at -80 °C. All subsequent expression studies were done with freshly grown bacterial cells that were isolated from LB/kanamycin plates streaked from the glycerol freezer stock.

BL21(DE3) *E. coli* cells containing either pE24CP1 or pET24c(+) (negative control) were streaked out onto LB/kanamycin plates from a glycerol freezer stock (above) and incubated overnight at 37 °C. One isolated colony was selected for inoculation of 2 mL of LB containing 30 µg/mL kanamycin in a 17 x 100 mm Falcon tube, and the culture was incubated at 37 °C with shaking (250 rpm) overnight. The overnight cultures were then used to inoculate 100 mL of fresh LB containing 30 µg/mL kanamycin. The new cultures were incubated at 37 °C with shaking until the A<sub>600</sub> reached between 0.4 and 0.6 absorbance units. One half of the pE24CP1 and pET24c(+) cultures were placed in new culture flasks and IPTG (isopropylthio-β-D-galactoside; Gibco BRL) was added to the new flasks to give a final concentration of 1 mM. The flasks were incubated an additional 3 h at 37 °C with shaking, and then the cells were harvested.

The harvested cells were centrifuged and the resulting cell pellet extracted by sonication (3 x 10 sec bursts) in 2 mL extraction buffer (50 mM (20 mM in the first experiment; Table 2) potassium phosphate buffer, pH 7.2, containing 0.14 M KCl, 0.32 mM reduced glutathione, 1% polyvinylpolypyrrolidone, and 0.1% Triton X 100 (0.01% lysozyme was included in the first experiment only)). The lysate represents the crude extracted enzyme after centrifugation at 17000 g for 10 min. In the first experiment (Table 2) a 20 to 60% ammonium sulfate precipitated enzyme fraction was also assayed. Solid ammonium sulfate was slowly added with stirring to 2 mL of the lysate to bring the concentration to 20% (w/v). After incubation on ice for approximately 15 min, the solution was centrifuged at 17000 g for 10 min. The supernatant liquid was harvested and solid ammonium sulfate was added to increase the concentration to 60% (w/v). After

centrifugation, the resulting pellet was resuspended in 1 mL of the extraction buffer.

A portion of the insoluble protein resulting from expression of *Arabidopsis* *p*-hydroxyphenylpyruvate dioxygenase in bacteria was utilized for N-terminal sequence analysis. The protein (approximately 180 µg) was suspended in 60 µL of extraction buffer and then diluted with 5 volumes of sample buffer (62.5 mM Tris, pH 6.8, 6 M urea, 160 mM dithiothreitol, 0.01% bromophenol blue) followed by intermittent vortexing for one hour at room temperature. A 1.5 mm thick, 12% polyacrylamide resolving gel was prepared for a Mini-Protein II dual slab cell (Bio-Rad, Hercules, CA) using the manufacturer's instructions. The polyacrylamide was allowed to polymerize for 3 h and then a stacking gel was prepared using a preparative comb. The running buffer was prepared according to the manufacturer's instructions with the addition of 0.1 mM sodium thioglycolate. The solubilized protein sample was electrophoretically separated using the manufacturer's instructions. When the bromophenol blue dye front reached the bottom of the gel, the gel was removed and equilibrated for 5 min in blotting buffer (10 mM CAPS, pH 11, 10% methanol, balance water). The gel was then placed in a Mini Trans-Blot Electrophoretic Transfer Cell (Bio-Rad), according to the manufacturer's instructions, with a ProBlott PVDF membrane (Applied Biosystems, Foster City, CA) treated according to the manufacturer's instruction. Electroblotting was done in the presence of blotting buffer at 50 volts for 45 min in an ice bath. The membrane was then rinsed in water and stained with Coomassie Blue as described in the ProBlott protocol. The major protein band was excised from the membrane and subjected to N-terminal amino acid sequencing on a Beckman (Fullerton, CA) LF3000 protein sequencer. The first 11 cycles identified S-K-F-V-R-K-N-P-K-S-D (see SEQ ID NO:3, amino acids 30-40), respectively. This is the expected N-terminus of the modified *Arabidopsis* *p*-hydroxyphenylpyruvate dioxygenase minus the initial methionine (amino acids 30-40, Figure 3).

30

### EXAMPLE 3

#### *p*-Hydroxyphenylpyruvate Dioxygenase Enzymatic Activity of the Plant Protein Expressed in *E. Coli*

Cell cultures with different plasmid constructs were extracted as described above and assayed by measuring the formation of  $^{14}\text{CO}_2$  from [1- $^{14}\text{C}$ ]-*p*-hydroxyphenylpyruvate or  $^{14}\text{CO}_2$  and  $^{14}\text{C}$ -homogentisate from [U- $^{14}\text{C}$ ]-*p*-hydroxyphenylpyruvate (Lindblad, B., (1971) *Clin. Chim. Acta* 34:113-121; and Lindstedt, S. and Odelhog, B., (1987) *Methods in Enzymology* 142:143-148). The labeled substrate was prepared from [1- $^{14}\text{C}$ ]-L-tyrosine

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(55 mCi/mmol; American Radiolabeled Chemicals, Inc., St. Louis, MO) or [U-<sup>14</sup>C]-L-tyrosine (498 mCi/mmol; DuPont NEN, Boston, MA). A 50-100 µL aliquot (5-10 µCi) of the of the labeled tyrosine stock solution was transferred to a 4 mL glass vial and blown to dryness in a stream of nitrogen at 45°C. To the vial  
5 was added 175 µL of 0.1 M phosphate buffer, pH 6.5, 5 µL catalase (28,700 units of C-100, Sigma Chemical Co., St. Louis, MO), and 20 µL L-amino acid oxidase (Sigma A-9253, 6.5 units/mL). The vial was then placed on a shaker water bath set at 30°C, 60 cycles/min, for 0.5 to 1 h. The reaction mix was then passed  
10 through a small column containing 400 µL Dowex AG 50W X8 cation exchange resin. The column was then washed with 1.5 mL of water and the eluant containing the labeled *p*-hydroxyphenylpyruvate was collected. The labeled substrate was either used immediately or stored at -80°C and used within a week after preparation.

The assay was performed in 14 mL culture tubes capped with serum  
15 stoppers through which a polypropylene well containing 200 µL of 1 N KOH was suspended. The reaction mixture contained 5,740 units of catalase, 100 µL of a freshly prepared 1:1 (v:v) mixture of 150 mM reduced glutathione and 3 mM dichlorophenolindophenol, 5 mM ascorbate, 0.1 mM ferrous sulfate (the ascorbate and ferrous sulfate were not present in the buffer used in the first experiment;  
20 Table 2), 50 µM unlabeled *p*-hydroxyphenylpyruvate, 1-25 µL of the enzyme extract, and 50 mM potassium phosphate buffer in a final volume of 980 µL. Unlabeled substrate was made fresh daily in 50 mM potassium phosphate buffer and allowed to equilibrate for at least 2 h at room temperature to insure that greater than 95% was in the keto form. The tubes were incubated for 10 min at  
25 30°C in a shaking water bath prior to adding 20 µL (0.04 µCi) of <sup>14</sup>C-*p*-hydroxyphenylpyruvate. The reaction was terminated after 60 min by injecting 500 µl of 1 N sulfuric acid through the serum stopper. The vials were left on the shaker for another 30 min to insure complete capture of the released <sup>14</sup>CO<sub>2</sub>. The serum caps were then removed and the wells cut and dropped into  
30 8 mL scintillation vials. Six mL of Formula-989 scintillation fluid (Packard Instruments, Meriden, CT) was added to the vials and the <sup>14</sup>C radioactivity was determined by scintillation counting. Table 2 summarizes the results of this experiment.

Table 2

*p*-Hydroxyphenylpyruvate Dioxygenase Activity of Extracts from  
*E. coli* Containing Different Plasmid Constructs

Plasmid	Inducer (1 mM IPTG)	Lysate		Ammonium Sulfate Precipitate	
		dpm * /mg	nmol/min x mg	dpm * /mg	nmol/min x mg
pET24c(+)	-	12,318	0.09	0	0.00
pET24c(+)	+	35,115	0.25	3,393	0.03
pE24CP1	-	24,607	0.17	126,761	0.89
pE24CP1	+	243,801	1.71	1,371,823	9.64

\*  $^{14}\text{C} : ^{12}\text{C} = 1 : 50$ ; sp. act. of  $^{14}\text{C}$ -*p*-hydroxyphenylpyruvate = 55 mCi/mmol

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The results show there was little or no *p*-hydroxyphenylpyruvate dioxygenase activity in any of the cell cultures that did not have the plasmid containing the nucleic acid fragment encoding *p*-hydroxyphenylpyruvate dioxygenase (pET24c(+)) and the inducer of gene expression (IPTG). The gene and inducer together resulted in a marked increase in activity.

10

In the experiment with  $[\text{U-}^{14}\text{C}]$  *p*-hydroxyphenylpyruvate ("HPPA"), where both  $^{14}\text{CO}_2$  and  $^{14}\text{C}$ -homogentisic acid were measured, the reaction was initiated by adding 50  $\mu\text{L}$  of labeled substrate (0.3  $\mu\text{Ci}$ ) and was terminated with 100  $\mu\text{L}$  of 10% phosphoric acid. The  $^{14}\text{CO}_2$  released was determined by scintillation counting, while the level of homogentisic acid was determined by HPLC on a Zorbax RX-C8 column (4.6 x 250 mm) with an in-line radioactivity detector. Aliquots of 1.7 to 15  $\mu\text{L}$  were taken from the reaction mix after centrifugation and diluted into the column equilibration buffer prior to injection. Separation was performed at ambient temperature with a flow rate of 1.0 mL/min and the following gradient with solvent A and B being water and methanol, each with 1% phosphoric acid: 0-2 min, isocratic at 95% A and 5% B; 2-17 min, linear gradient from 95 to 75% A and 5 to 25% B; 17-19 min linear gradient from 75 to 5% A and 25 to 95% B; 19-22 min, isocratic at 5% A and 95% B; 22-24 min, linear gradient from 5% to 95% A and 95 to 5% B. In this system homogentisate eluted at 10.8 min. The results from this experiment are shown in Table 3.

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20

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Table 3

*p*-Hydroxyphenylpyruvate Dioxygenase Activity of Cell Extracts  
Determined by CO<sub>2</sub> Release and Homogentisic Acid Synthesis  
from [U-<sup>14</sup>C] *p*-Hydroxyphenylpyruvate

Plasmid	Inducer (1 mM IPTG)	nmol/min x mg*	
		<sup>14</sup> CO <sub>2</sub>	Homogentisic acid
pET24c(+)	-	0.00	0.00
pET24c(+)	+	0.19	0.00
pE24CP1	-	4.68	4.76
pE24CP1	+	29.12	29.82

\* <sup>14</sup>C : <sup>12</sup>C = 1 : 87.7; sp. act. of <sup>14</sup>C[U]-*p*-HPPA = 498 mCi/mmol

There was a tight correlation between the results from the assays of the two products of the reaction. The results confirmed there was no significant *p*-hydroxyphenylpyruvate dioxygenase activity in either cell culture that did not contain the nucleic acid fragment encoding *p*-hydroxyphenylpyruvate dioxygenase. There was measureable enzyme activity in the absence of the inducer, but when the inducer was added the activity increased greater than six-fold over uninduced cultures. These results and those of Table 2 clearly show that the nucleic acid fragment isolated and overexpressed in *E. coli* cells encodes a protein that catalyzes the conversion of *p*-hydroxyphenylpyruvate to homogentisate with the release of CO<sub>2</sub>.

The overexpressed protein was also assayed spectrophotometrically at ambient temperature using the enol borate-tautomerase assay (Lin, E. C. C. et al., (1958) *J. Biol. Chem.* 233:668-673). The assay buffer contained 0.4 M borate (adjusted to pH 7.2 with 0.2 M sodium borate), 4 mM ascorbate, 2.5 mM EDTA, 40 μM *p*-hydroxyphenylpyruvate, and 0.5 units of tautomerase (Sigma T-6004) per 10 mL buffer. The reaction mix was used when the tautomerization of the substrate was complete (when absorbance at 308 nm had stabilized). The assay was initiated by adding 40 μL of the cell extracts to 960 μL of the assay buffer, and the reaction was followed by measuring the decrease in absorbance at 308 nm. Table 4 summarizes the results with extracts of the same four cell cultures described in Table 3.

Table 4  
Spectrophotometric Assay of *p*-Hydroxyphenylpyruvate  
Dioxygenase Activity of Cell Extracts

Plasmid	Inducer (1 mM IPTG)	nmol <i>p</i> -HP lost/min x mg*
pET24c(+)	-	1.58
pET24c(+)	+	2.73
pE24CP1	-	4.91
pE24CP1	+	22.32

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\* Loss of *p*-hydroxyphenylpyruvate based on a molar extinction coefficient for the equilibrium mixture of 9850 as reported by Lin et al. ((1958) *J. Biol. Chem.* 233: 668-673).

#### EXAMPLE 4

##### 10 Inhibition of *p*-Hydroxyphenylpyruvate Dioxygenase by Commercial Herbicides

The enzymatic activity of the overexpressed protein is inhibited by two herbicides known to inhibit plant *p*-hydroxyphenylpyruvate dioxygenase: Sulcotrione (2-(2-chloro-4-methanesulfonylbenzoyl)-1,3-cyclohexanedione); and Isoxaflutole (5-cyclopropylisoxazol-4-yl 2-mesyl-4-trifluoromethylphenyl ketone). These two compounds were tested against the overexpressed protein using both the  $^{14}\text{CO}_2$  and the continuous spectrophotometric enol borate-  
15 tautomerase assays. Both compounds were added to the assay buffers in 10  $\mu\text{L}$  of acetone or dimethyl sulfoxide. The  $I_{50}$  values (concentration inhibiting the enzyme 50%) were calculated based on the percent inhibition observed over  
20 several concentrations of the inhibitor. The results of the assays are shown in Table 5.

Table 5

$I_{50}$  Values of Inhibitors of Plant *p*-Hydroxyphenylpyruvate Dioxygenase

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Compound	$I_{50}$ value (nM) derived from	
	$^{14}\text{CO}_2$ assay	spectrophotometric assay
sulcotrione	43	44
isoxaflutole	409	1042

These results clearly show that the *p*-hydroxyphenylpyruvate dioxygenase activity of the overexpressed protein is inhibited by commercial herbicides that have inhibition of this enzyme as their mode of action. Moreover, the continuous  
30 spectrophotometric assay gave similar  $I_{50}$  values to those obtained with the  $^{14}\text{CO}_2$  assay. The spectrophotometric assay can be adapted to a high capacity screen for

inhibitors of *p*-hydroxyphenylpyruvate dioxygenase by adapting it to a microtiter plate assay combined with a plate reader that would read at or near 308 nm.

Furthermore, any colorimetric or fluorescent assay for homogentisate or

*p*-hydroxyphenylpyruvate would also be able to be readily adapted into a high

- 5 capacity screen for inhibitors of this enzyme. The isolated overexpressed enzyme has sufficient activity to be used directly in a spectrophotometric assay or it can be further purified for enhanced assay sensitivity.

#### EXAMPLE 5

##### Re-construction of the Full-length *p*-Hydroxyphenylpyruvate Dioxygenase Gene 10 for Production of Active, Stable Enzyme in Bacteria

- The plasmid pT7BlueR+PDO2, described in Example 2 and containing the full-length *p*-hydroxyphenylpyruvate dioxygenase gene, proved to have incorrect sequence at the EcoRI site. This was re-sequenced so that an oligonucleotide could be designed to replace the EcoRI site with an NdeI site using conventional
- 15 loop-out mutagenesis. The oligonucleotide was designed so that this procedure also introduced an ATG initiation codon at the 5'- end of the *p*-hydroxyphenylpyruvate dioxygenase gene followed by the full-length *p*-hydroxyphenylpyruvate dioxygenase sequence. After mutagenesis, the clone was amplified in *E. coli* and the plasmid was purified. The resulting full-length gene, "PDO-B", was then
- 20 digested with the enzymes using NdeI and NheI, and the ~820 bp fragment used to replace the NdeI - Nhe I segment of the truncated *p*-hydroxyphenylpyruvate dioxygenase gene, "PDO-A," in pE24CP1 (Example 1). The resulting plasmid, pE24PDO-B can be expressed in bacteria to produce the full-length *Arabidopsis* *p*-hydroxyphenylpyruvate dioxygenase enzyme as determined by enzyme activity
- 25 and N-terminal sequence analysis.

#### EXAMPLE 6

##### Enhanced Stability of Full Length Construct Over the Truncated Construct

- Two different constructs for *Arabidopsis thaliana* *p*-hydroxyphenylpyruvate dioxygenase, one containing the full-length sequence, PDO-B as
- 30 described in Example 5 and produced from plasmid pE24PDO-B, and one containing the truncated sequence lacking the putative chloroplast leader sequence, PDO-A as produced from plasmid pE24CP1, were both purified to the same extent using a Pharmacia phenyl Sepharose hydrophobic interaction column followed by gel filtration chromatography on Pharmacia Sephacryl 300. The two
- 35 proteins were diluted to 1 mg/mL in 20 mM bis tris-propane buffer, pH 7.2 containing 5 mM ascorbate, 1 mM reduced glutathione and 0.1 mM ferrous ammonium sulfate and stored in a refrigerator at 4 °C for up to 10 days. Aliquots were removed at various times and assayed for activity using the tautomerase



coupled spectrophotometric assay. Under these conditions the half-life for the activity of the full length enzyme was 4 days, whereas the truncated enzyme preparation had a half-life of 9 to 10 hours. In addition, the activity of the full length enzyme could be restored by incubation with iron and reducing agent, reduced glutathione or ascorbate, or by dialysis against buffer containing iron and reducing agent. In contrast, the activity of the truncated enzyme could not be restored by incubation with or dialysis against buffer containing iron and reducing agent. The full-length enzyme was also more stable in the spectrophotometric assay showing a 2 to 3 times longer useful linear region than the truncated enzyme. Both enzyme preparations showed similar  $I_{50}$  values with the herbicidally active inhibitors.

These results clearly show that the full-length PDO-B construct has decided advantages over the truncated enzyme due to the enhanced stability under storage conditions, in the spectrophotometric assay and in the reversible reconstitution of activity in the presence of iron and reducing agent. While both enzyme constructs can be used for screening of inhibitors, the PDO-B enzyme is preferred for this application and is far superior for mechanistic and structural studies.

#### EXAMPLE 7

##### 20 Cloning of the Maize *p*-Hydroxyphenylpyruvate Dioxygenase Gene

Approximately 600,000 plaques of a Stratagene maize Uni-Zap cDNA library (from young plants) were screened by filter hybridization under moderate stringency using a heterologous probe. The probe was prepared by PCR and was a 916 bp fragment of DNA having the sequence defined by the region extending from position 263 to 1178 of SEQ ID NO:14. Twenty-four positive phage clones were identified in the primary screen, and eleven phage clones were recovered from a secondary screen. Seven positive clones were submitted for sequencing, and four showed significant conservation sequence at the amino acid level when compared with the *Arabidopsis thaliana* *p*-hydroxyphenylpyruvate dioxygenase protein. The longest of the four contained an insert of 988 bp and showed 70% identity and 78% similarity with the *Arabidopsis* protein, but was lacking approximately 550 bp corresponding to the amino terminal end of the protein.

Attempts to obtain a full-length cDNA of the maize *p*-hydroxyphenylpyruvate dioxygenase gene were unsuccessful, possibly because the secondary structure of the RNA inhibited efficient reverse transcription of this transcript. Two additional cDNA libraries were screened and clones long enough to contain a full-length cDNA were sequenced. All of these clones were shown to be chimeras. Therefore a genomic library was screened to obtain the 5' one-third of

the gene. Approximately 1 million clones from a Clontech *Zea mays* (var. B73) library in the phage vector EMBL3 (whole seedlings, 2 leaf stage) were screened using a 415 bp EcoRI-BssHII fragment containing the 5' end of the truncated corn *p*-hydroxyphenylpyruvate dioxygenase cDNA (clone H1011C). Eight positive  
5 primary phage clones were plated and screened, and four secondary clones were picked. DNA was prepared from each using the Qiagen Lambda midi-kit. Restriction digests with Sall or EcoRI indicated that two clones were the same. DNA samples from the remaining 3 clones (11.1.3, 13.1.1, and 21.2.1) were digested with Sall, EcoRI, or Sall and EcoRI, prepared for Southern analysis, and  
10 probed with the full length *Arabidopsis p*-hydroxyphenylpyruvate dioxygenase gene. Two of the clones (11.1.3 and 13.1.1) showed sequence conservation, and these homologous fragments were subcloned and sequenced. Both clones appeared to contain the full-length gene and each contained one intron near the 3' end of the gene. However, there were differences between the sequences of the  
15 two clones indicating that they may be two different genes or one may be a pseudogene. The sequence of clone 11.1.3 matched the cDNA sequence, and this clone was used to construct a full length *p*-hydroxyphenylpyruvate dioxygenase coding region.

The gene was contained on two adjacent fragments, a 3.5 kb EcoRI - Sall  
20 fragment and a 2 kb Sall fragment. Both were subcloned into pBluescript SKII+ resulting in the plasmids pES1113 and pSal1113. pES1113 was digested with SpeI to release approximately 2.7 kb of upstream sequence and then religated, resulting in a plasmid with an insert of 747 base pairs (pSPE1). pSPE1 was digested with Sall to linearize the plasmid and ligated with the 2 kb Sall fragment  
25 from pSal1113, which had been released by digestion with Sall and gel purified. Orientation was confirmed by digestion with SpeI and Bpu1102I and the correct plasmid was named p1113. In order to remove the intron contained in the 3' end of the genomic clone, the plasmid was digested with Bpu1102I and XhoI and the 3.9 kb fragment containing the vector and 5' part of the gene was gel purified.  
30 The corresponding 882 bp Bpu1102I-XhoI fragment from pH1011c (cDNA) was gel purified and ligated with this 3.9 kb fragment resulting in the clone pMPDO (ATCC 209120), which contains a 1782 bp insert. There are 260 base pairs upstream of the putative ATG and 189 base pairs downstream of the stop codon. The full-length sequence was confirmed by sequencing across the insert. The  
35 nucleic acid sequence and the deduced protein sequence for corn *p*-hydroxyphenylpyruvate dioxygenase are presented in SEQ ID NOS:10 and 11, respectively. The sequences for *p*-hydroxyphenylpyruvate dioxygenases obtained from corn and *Arabidopsis* were compared using the "Gap" program of GCG

(Program Manual for the Wisconsin Package, Version 9.0-OpenVMS, December 1996, Genetics Computer Group, 575 Science Drive, Madison, WI, USA 53711). The results of these comparisons indicated that these functions are approximately 67% identical at the nucleotide level, and they possess 69% similarity and 62% identity at the amino acid level. The predicted amino acid sequence of corn *p*-hydroxyphenylpyruvate dioxygenase is compared with that from *Arabidopsis* and other eukaryotes in Figure 3.

#### EXAMPLE 8

##### Composition of a cDNA Library; Isolation and Sequencing of cDNA Clones

10 A cDNA library representing mRNAs from developing seeds of *Vernonia galamenensis* that had just begun production of vernolic acid was prepared. The library was prepared in a Uni-ZAP™ XR vector according to the manufacturer's protocol (Stratagene Cloning Systems, La Jolla, CA). Conversion of the Uni-ZAP™ XR library into a plasmid library was accomplished according to the  
15 protocol provided by Stratagene. Upon conversion, cDNA inserts were contained in the plasmid vector pBluescript. cDNA inserts from randomly picked bacterial colonies containing recombinant pBluescript plasmids were amplified via polymerase chain reaction using primers specific for vector sequences flanking the inserted cDNA sequences. Amplified insert DNAs were sequenced in dye-  
20 primer sequencing reactions to generate partial cDNA sequences (expressed sequence tags or "ESTs"; see Adams, M. D. et al., (1991) *Science* 252:1651). The resulting ESTs were analyzed using a Perkin Elmer Model 377 fluorescent sequencer.

#### EXAMPLE 9

##### Identification and Characterization of cDNA Clones

25 ESTs encoding *Vernonia galamenensis* enzymes were identified by conducting BLAST (Basic Local Alignment Search Tool; Altschul, S. F. et al., (1993) *J. Mol. Biol.* 215:403-410; see also [www.ncbi.nlm.nih.gov/BLAST/](http://www.ncbi.nlm.nih.gov/BLAST/)) searches for similarity to sequences contained in the BLAST "nr" database  
30 (comprising all non-redundant GenBank CDS translations, sequences derived from the 3-dimensional structure Brookhaven Protein Data Bank, the last major release of the SWISS-PROT protein sequence database, EMBL, and DDBJ databases). The cDNA sequences obtained in Example 9 were analyzed for similarity to all publicly available DNA sequences contained in the "nr" database  
35 using the BLASTN algorithm provided by the National Center for Biotechnology Information (NCBI). The DNA sequences were translated in all reading frames and compared for similarity to all publicly available protein sequences contained in the "nr" database using the BLASTX algorithm (Gish, W. and States, D. J.

(1993) *Nature Genetics* 3:266-272) provided by the NCBI. For convenience, the P-value (probability) of observing a match of a cDNA sequence to a sequence contained in the searched databases merely by chance as calculated by BLAST are reported herein as "pLog" values, which represent the negative of the logarithm of the reported P-value. Accordingly, the greater the pLog value, the greater the likelihood that the cDNA sequence and the BLAST "hit" represent homologous proteins.

The BLASTX search using clone vs1.pk0015.b2 revealed similarity of the protein encoded by the cDNA to a number of *p*-hydroxyphenylpyruvate dioxygenases from sources other than plants. The three most similar *p*-hydroxyphenylpyruvate dioxygenase proteins were a streptomycete *p*-hydroxyphenylpyruvate dioxygenase (GenBank Accession No. U11864; pLog = 8.34), a rat *p*-hydroxyphenylpyruvate dioxygenase (GenBank Accession No. M18405; pLog = 7.66), and a human *p*-hydroxyphenylpyruvate dioxygenase (GenBank Accession No. U29895; pLog = 7.60). SEQ ID NO:16 shows the nucleotide sequence of a portion of the *Vernonia galamenensis* cDNA in clone vs1.pk0015.b2. Sequence alignments and BLAST scores and probabilities indicate that the instant nucleic acid fragment encodes a portion of *Vernonia galamenensis* *p*-hydroxyphenylpyruvate dioxygenase.

20

SEQUENCE LISTING

## (1) GENERAL INFORMATION:

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- (ii) TITLE OF INVENTION: PLANT GENE FOR *p*-HYDROXY-PHENYLPYRUVATE DIOXYGENASE
- (iii) NUMBER OF SEQUENCES: 16
- (iv) COMPUTER READABLE FORM:
  - (A) MEDIUM TYPE: DISKETTE, 3.50 INCH
  - (B) COMPUTER: IBM PC COMPATIBLE
  - (C) OPERATING SYSTEM: MICROSOFT WORD FOR WINDOWS 95
  - (D) SOFTWARE: MICROSOFT WORD VERSION 7.0A
- (v) CURRENT APPLICATION DATA:
  - (A) APPLICATION NUMBER:
  - (B) FILING DATE:
  - (C) CLASSIFICATION:
- (vi) PRIOR APPLICATION DATA:
  - (A) APPLICATION NUMBER: 60/021,364
  - (B) FILING DATE: JUNE 27, 1996
- (vii) ATTORNEY/AGENT INFORMATION:
  - (A) NAME: FLOYD, LINDA AXAMETHY
  - (B) REGISTRATION NUMBER: 33,692
  - (C) REFERENCE/DOCKET NUMBER: BA-9120

## (2) INFORMATION FOR SEQ ID NO:1:

- (i) SEQUENCE CHARACTERISTICS:  
 (A) LENGTH: 233 base pairs  
 (B) TYPE: nucleic acid  
 (C) STRANDEDNESS: single  
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

## (xi) SEQUENCE DESCRIPTION: SEQ ID NO:1:

CAAGAAACGN GTCGNCGACG TGCTCAGCGA TGATCAGATC AAGGAGTGTG AGGAATTAGG 60  
 GATTCTNTA GACAGAGATG ATCAAGGGAC GTTNCCTCAA ATCTNCACAA AACCACTAGG 120  
 TGACAGGCCG ACGNTATTTA TAGAGATAAT CCAGAGNGTA GGATGCATGA TGAAAGATGT 180  
 GGAAGGGANG GCTTACCAGA GTGGAGNATN TNGTGGTTTT GGCAAAGGCA ATT 233

## (2) INFORMATION FOR SEQ ID NO:2:

- (i) SEQUENCE CHARACTERISTICS:  
 (A) LENGTH: 1448 base pairs  
 (B) TYPE: nucleic acid  
 (C) STRANDEDNESS: single  
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

- (ix) FEATURE:  
 (A) NAME/KEY: CDS  
 (B) LOCATION: 9..1343

## (xi) SEQUENCE DESCRIPTION: SEQ ID NO:2:

TGAAATCA ATG GGC CAC CAA AAC GCC GCC GTT TCA GAG AAT CAA AAC CAT 50  
 Met Gly His Gln Asn Ala Ala Val Ser Glu Asn Gln Asn His  
 1 5 10

GAT GAC GGC GCT GCG TCG TCG CCG GGA TTC AAG CTC GTC GGA TTT TCC 98  
 Asp Asp Gly Ala Ala Ser Ser Pro Gly Phe Lys Leu Val Gly Phe Ser  
 15 20 25 30

AAG TTC GTA AGA AAG AAT CCA AAG TCT GAT AAA TTC AAG GTT AAG CGC 146  
 Lys Phe Val Arg Lys Asn Pro Lys Ser Asp Lys Phe Lys Val Lys Arg  
 35 40 45

TTC CAT CAC ATC GAG TTC TGG TGC GGG GAC GCA ACC AAC GTC GCT CGT 194  
 Phe His His Ile Glu Phe Trp Cys Gly Asp Ala Thr Asn Val Ala Arg  
 50 55 60

CGC TTC TCC TGG GGT CTG GGG ATG AGA TTC TCC GCC AAA TCC GAT CTT 242  
 Arg Phe Ser Trp Gly Leu Gly Met Arg Phe Ser Ala Lys Ser Asp Leu  
 65 70 75

TCC ACC GGA AAC ATG GTT CAC GCC TCT TAC CTA CTC ACC TCC GGT GAA 290  
 Ser Thr Gly Asn Met Val His Ala Ser Tyr Leu Leu Thr Ser Gly Glu  
 80 85 90

CTC CGA TTC CTT TTC ACT GCT CCT TAC TCT CCG TCT CTC TCC GGC GGA 338  
 Leu Arg Phe Leu Phe Thr Ala Pro Tyr Ser Pro Ser Leu Ser Gly Gly  
 95 100 105 110

GAG ATT AAA CCG ACA ACC ACA GGT TCT ATC CCA AGT TTC GAT CAC GGG Glu Ile Lys Pro Thr Thr Thr Gly Ser Ile Pro Ser Phe Asp His Gly 115 120 125	386
TCT TGT CGG TCC TTC TTC TCT TCA CAT GGT CTC GGT GTT AGA CCC GTT Ser Cys Arg Ser Phe Phe Ser Ser His Gly Leu Gly Val Arg Pro Val 130 135 140	434
GCG ATT GAA GTA GAA GAC GCG GAG TCA GCT TTC TCC ATC AGT GTA GCT Ala Ile Glu Val Glu Asp Ala Glu Ser Ala Phe Ser Ile Ser Val Ala 145 150 155	482
AAT GGC GCT ATT CCT TCG TCG CCT CCT ATC GTC CTC AAT GAA GCA GTT Asn Gly Ala Ile Pro Ser Ser Pro Pro Ile Val Leu Asn Glu Ala Val 160 165 170	530
ACG ATC GCT GAG GTT AAA CTA TAC GGC GAT GTT GTT CTC CGA TAT GTT Thr Ile Ala Glu Val Lys Leu Tyr Gly Asp Val Val Leu Arg Tyr Val 175 180 185 190	578
AGT TAC AAA GCA GAA GAT ACC GAA AAA TCC GAA TTC TTG CCA GGG TTC Ser Tyr Lys Ala Glu Asp Thr Glu Lys Ser Glu Phe Leu Pro Gly Phe 195 200 205	626
GAG CGT GTA GAG GAT GCG TCG TCG TTC CCA TTG GAT TAT GGT ATC CGG Glu Arg Val Glu Asp Ala Ser Ser Phe Pro Leu Asp Tyr Gly Ile Arg 210 215 220	674
CGG CTT GAC CAC GCC GTG GGA AAC GTT CCT GAG CTT GGT CCG GCT TTA Arg Leu Asp His Ala Val Gly Asn Val Pro Glu Leu Gly Pro Ala Leu 225 230 235	722
ACT TAT GTA GCG GGG TTC ACT GGT TTT CAC CAA TTC GCA GAG TTC ACA Thr Tyr Val Ala Gly Phe Thr Gly Phe His Gln Phe Ala Glu Phe Thr 240 245 250	770
GCA GAC GAC GTT GGA ACC GCC GAG AGC GGT TTA AAT TCA GCG GTC CTG Ala Asp Asp Val Gly Thr Ala Glu Ser Gly Leu Asn Ser Ala Val Leu 255 260 265 270	818
GCT AGC AAT GAT GAA ATG GTT CTT CTA CCG ATT AAC GAG CCA GTG CAC Ala Ser Asn Asp Glu Met Val Leu Leu Pro Ile Asn Glu Pro Val His 275 280 285	866
GGA ACA AAG AGG AAG AGT CAG ATT CAG ACG TAT TTG GAA CAT AAC GAA Gly Thr Lys Arg Lys Ser Gln Ile Gln Thr Tyr Leu Glu His Asn Glu 290 295 300	914
GGC GCA GGG CTA CAA CAT CTG GCT CTG ATG AGT GAA GAC ATA TTC AGG Gly Ala Gly Leu Gln His Leu Ala Leu Met Ser Glu Asp Ile Phe Arg 305 310 315	962
ACC CTG AGA GAG ATG AGG AAG AGG AGC AGT ATT GGA GGA TTC GAC TTC Thr Leu Arg Glu Met Arg Lys Arg Ser Ser Ile Gly Gly Phe Asp Phe 320 325 330	1010
ATG CCT TCT CCT CCG CCT ACT TAC TAC CAG AAT CTC AAG AAA CGG GTC Met Pro Ser Pro Pro Pro Thr Tyr Tyr Gln Asn Leu Lys Lys Arg Val 335 340 345 350	1058
GGC GAC GTG CTC AGC GAT GAT CAG ATC AAG GAG TGT GAG GAA TTA GGG Gly Asp Val Leu Ser Asp Asp Gln Ile Lys Glu Cys Glu Glu Leu Gly 355 360 365	1106

ATT CTT GTA GAC AGA GAT GAT CAA GGG ACG TTG CTT CAA ATC TTC ACA 1154  
 Ile Leu Val Asp Arg Asp Asp Gln Gly Thr Leu Leu Gln Ile Phe Thr  
 370 375 380

AAA CCA CTA GGT GAC AGG CCG ACG ATA TTT ATA GAG ATA ATC CAG AGA 1202  
 Lys Pro Leu Gly Asp Arg Pro Thr Ile Phe Ile Glu Ile Ile Gln Arg  
 385 390 395

GTA GGA TGC ATG ATG AAA GAT GAG GAA GGG AAG GCT TAC CAG AGT GGA 1250  
 Val Gly Cys Met Met Lys Asp Glu Gly Lys Ala Tyr Gln Ser Gly  
 400 405 410

GGA TGT GGT GGT TTT GCC AAA GGC AAT TTC TCT GAG CTC TTC AAG TCC 1298  
 Gly Cys Gly Gly Phe Ala Lys Gly Asn Phe Ser Glu Leu Phe Lys Ser  
 415 420 425 430

ATT GAA GAA TAC GAA AAG ACT CTT GAA GCC AAA CAG TTA GTG GGA 1343  
 Ile Glu Glu Tyr Glu Lys Thr Leu Glu Ala Lys Gln Leu Val Gly  
 435 440 445

TGAACAAGAA GAAGAACCAA CTAAAGGATT GTGTAATTAA TGTAAGAACTG TTTTATCTTA 1403

TCAAAACAAT GTATACAACA TCTCATTTAA AAACGAGATC AATCC 1448

## (2) INFORMATION FOR SEQ ID NO:3:

## (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 445 amino acids
- (B) TYPE: amino acid
- (D) TOPOLOGY: linear

## (ii) MOLECULE TYPE: protein

## (xi) SEQUENCE DESCRIPTION: SEQ ID NO:3:

Met Gly His Gln Asn Ala Ala Val Ser Glu Asn Gln Asn His Asp Asp  
 1 5 10 15

Gly Ala Ala Ser Ser Pro Gly Phe Lys Leu Val Gly Phe Ser Lys Phe  
 20 25 30

Val Arg Lys Asn Pro Lys Ser Asp Lys Phe Lys Val Lys Arg Phe His  
 35 40 45

His Ile Glu Phe Trp Cys Gly Asp Ala Thr Asn Val Ala Arg Arg Phe  
 50 55 60

Ser Trp Gly Leu Gly Met Arg Phe Ser Ala Lys Ser Asp Leu Ser Thr  
 65 70 75 80

Gly Asn Met Val His Ala Ser Tyr Leu Leu Thr Ser Gly Glu Leu Arg  
 85 90 95

Phe Leu Phe Thr Ala Pro Tyr Ser Pro Ser Leu Ser Gly Gly Glu Ile  
 100 105 110

Lys Pro Thr Thr Thr Gly Ser Ile Pro Ser Phe Asp His Gly Ser Cys  
 115 120 125

Arg Ser Phe Phe Ser Ser His Gly Leu Gly Val Arg Pro Val Ala Ile  
 130 135 140

Glu Val Glu Asp Ala Glu Ser Ala Phe Ser Ile Ser Val Ala Asn Gly  
 145 150 155 160



Ala Ile Pro Ser Ser Pro Pro Ile Val Leu Asn Glu Ala Val Thr Ile  
165 170 175

Ala Glu Val Lys Leu Tyr Gly Asp Val Val Leu Arg Tyr Val Ser Tyr  
180 185 190

Lys Ala Glu Asp Thr Glu Lys Ser Glu Phe Leu Pro Gly Phe Glu Arg  
195 200 205

Val Glu Asp Ala Ser Ser Phe Pro Leu Asp Tyr Gly Ile Arg Arg Leu  
210 215 220

Asp His Ala Val Gly Asn Val Pro Glu Leu Gly Pro Ala Leu Thr Tyr  
225 230 235 240

Val Ala Gly Phe Thr Gly Phe His Gln Phe Ala Glu Phe Thr Ala Asp  
245 250 255

Asp Val Gly Thr Ala Glu Ser Gly Leu Asn Ser Ala Val Leu Ala Ser  
260 265 270

Asn Asp Glu Met Val Leu Leu Pro Ile Asn Glu Pro Val His Gly Thr  
275 280 285

Lys Arg Lys Ser Gln Ile Gln Thr Tyr Leu Glu His Asn Glu Gly Ala  
290 295 300

Gly Leu Gln His Leu Ala Leu Met Ser Glu Asp Ile Phe Arg Thr Leu  
305 310 315 320

Arg Glu Met Arg Lys Arg Ser Ser Ile Gly Gly Phe Asp Phe Met Pro  
325 330 335

Ser Pro Pro Pro Thr Tyr Tyr Gln Asn Leu Lys Lys Arg Val Gly Asp  
340 345 350

Val Leu Ser Asp Asp Gln Ile Lys Glu Cys Glu Glu Leu Gly Ile Leu  
355 360 365

Val Asp Arg Asp Asp Gln Gly Thr Leu Leu Gln Ile Phe Thr Lys Pro  
370 375 380

Leu Gly Asp Arg Pro Thr Ile Phe Ile Glu Ile Ile Gln Arg Val Gly  
385 390 395 400

Cys Met Met Lys Asp Glu Glu Gly Lys Ala Tyr Gln Ser Gly Gly Cys  
405 410 415

Gly Gly Phe Ala Lys Gly Asn Phe Ser Glu Leu Phe Lys Ser Ile Glu  
420 425 430

Glu Tyr Glu Lys Thr Leu Glu Ala Lys Gln Leu Val Gly  
435 440 445

## (2) INFORMATION FOR SEQ ID NO:4:

## (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 53 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

## (ii) MOLECULE TYPE: DNA (genomic)

## (xi) SEQUENCE DESCRIPTION: SEQ ID NO:4:

TATGTCCAAG TTCGTAAGAA AGAATCCAAA GTCTGATAAA TTCAAGGTTA AGC 53

## (2) INFORMATION FOR SEQ ID NO:5:

## (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 51 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

## (ii) MOLECULE TYPE: DNA (genomic)

## (xi) SEQUENCE DESCRIPTION: SEQ ID NO:5:

GCTTAACCTT GAATTTATCA GACTTTGGAT TCTTTCTTAC GAACTTGGAC A 51

## (2) INFORMATION FOR SEQ ID NO:6:

## (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 392 amino acids
- (B) TYPE: amino acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

## (ii) MOLECULE TYPE: protein

## (xi) SEQUENCE DESCRIPTION: SEQ ID NO:6:

Thr Ser Tyr Ser Asp Lys Gly Glu Lys Pro Glu Arg Gly Arg Phe Leu  
 1 5 10 15  
 His Phe His Ser Val Thr Phe Trp Val Gly Asn Ala Lys Gln Ala Ala  
 20 25 30  
 Ser Tyr Tyr Cys Ser Lys Ile Gly Phe Glu Pro Leu Ala Tyr Lys Gly  
 35 40 45  
 Leu Glu Thr Gly Ser Arg Glu Val Val Ser His Val Val Lys Gln Asp  
 50 55 60  
 Lys Ile Val Phe Val Phe Ser Ser Ala Leu Asn Pro Trp Asn Lys Glu  
 65 70 75 80  
 Met Gly Asp His Leu Val Lys His Gly Asp Gly Val Lys Asp Ile Ala  
 85 90 95  
 Phe Glu Val Glu Asp Cys Asp Tyr Ile Val Gln Lys Ala Arg Glu Arg  
 100 105 110  
 Gly Ala Ile Ile Val Arg Glu Glu Val Cys Cys Ala Ala Asp Val Arg  
 115 120 125  
 Gly His His Thr Pro Leu Asp Arg Ala Arg Gln Val Trp Glu Gly Thr  
 130 135 140  
 Leu Val Glu Lys Met Thr Phe Cys Leu Asp Ser Arg Pro Gln Pro Ser  
 145 150 155 160  
 Gln Thr Leu Leu His Arg Leu Leu Leu Ser Lys Leu Pro Lys Cys Gly  
 165 170 175  
 Leu Glu Ile Ile Asp His Ile Val Gly Asn Gln Pro Asp Gln Glu Met  
 180 185 190

Glu Ser Ala Ser Gln Trp Tyr Met Arg Asn Leu Gln Phe His Arg Phe  
 195 200 205  
 Trp Ser Val Asp Asp Thr Gln Ile His Thr Glu Tyr Ser Ala Leu Arg  
 210 215 220  
 Ser Val Val Met Ala Asn Tyr Glu Glu Ser Ile Lys Met Pro Ile Asn  
 225 230 235 240  
 Glu Pro Ala Pro Gly Lys Lys Lys Ser Gln Ile Gln Glu Tyr Val Asp  
 245 250 255  
 Tyr Asn Gly Gly Ala Gly Val Gln His Ile Ala Leu Lys Thr Glu Asp  
 260 265 270  
 Ile Ile Thr Ala Ile Arg Ser Leu Arg Glu Arg Gly Val Glu Phe Leu  
 275 280 285  
 Ala Val Pro Phe Thr Tyr Tyr Lys Gln Leu Gln Glu Lys Leu Lys Ser  
 290 295 300  
 Ala Lys Ile Arg Val Lys Glu Ser Ile Asp Val Leu Glu Glu Leu Lys  
 305 310 315 320  
 Ile Leu Val Asp Tyr Asp Glu Lys Gly Tyr Leu Leu Gln Ile Phe Thr  
 325 330 335  
 Lys Pro Met Gln Asp Arg Pro Thr Val Phe Leu Glu Val Ile Gln Arg  
 340 345 350  
 Asn Asn His Gln Gly Phe Gly Ala Gly Asn Phe Asn Ser Leu Phe Lys  
 355 360 365  
 Ala Phe Glu Glu Glu Gln Glu Leu Arg Gly Asn Leu Thr Asp Thr Asp  
 370 375 380  
 Pro Asn Gly Val Pro Phe Arg Leu  
 385 390

## (2) INFORMATION FOR SEQ ID NO:7:

## (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 392 amino acids  
 (B) TYPE: amino acid  
 (C) STRANDEDNESS: single  
 (D) TOPOLOGY: linear

## (ii) MOLECULE TYPE: protein

## (xi) SEQUENCE DESCRIPTION: SEQ ID NO:7:

Thr Ser Tyr Ser Asp Lys Gly Glu Lys Pro Glu Arg Gly Arg Phe Leu  
 1 5 10 15  
 His Phe His Ser Val Thr Phe Trp Val Gly Asn Ala Lys Gln Ala Ala  
 20 25 30  
 Ser Tyr Tyr Cys Ser Lys Ile Gly Phe Glu Pro Leu Ala Tyr Lys Gly  
 35 40 45  
 Leu Glu Thr Gly Ser Arg Glu Val Val Ser His Val Val Lys Gln Asp  
 50 55 60  
 Lys Ile Val Phe Val Phe Ser Ser Ala Leu Asn Pro Trp Asn Lys Glu  
 65 70 75 80

Met Gly Asp His Leu Val Lys His Gly Asp Gly Val Lys Asp Ile Ala  
 85 90 95  
 Phe Glu Val Glu Asp Cys Asp Tyr Ile Val Gln Lys Ala Arg Glu Arg  
 100 105 110  
 Gly Ala Ile Ile Val Arg Glu Glu Val Cys Cys Ala Ala Asp Val Arg  
 115 120 125  
 Gly His His Thr Pro Leu Asp Arg Ala Arg Gln Val Trp Glu Gly Thr  
 130 135 140  
 Leu Val Glu Lys Met Thr Phe Cys Leu Asp Ser Arg Pro Gln Pro Ser  
 145 150 155 160  
 Gln Thr Leu Leu His Arg Leu Leu Leu Ser Lys Leu Pro Lys Cys Gly  
 165 170 175  
 Leu Glu Ile Ile Asp His Ile Val Gly Asn Gln Pro Asp Gln Glu Met  
 180 185 190  
 Glu Ser Ala Ser Gln Trp Tyr Met Arg Asn Leu Gln Phe His Arg Phe  
 195 200 205  
 Trp Ser Val Asp Asp Thr Gln Ile His Thr Glu Tyr Ser Ala Leu Arg  
 210 215 220  
 Ser Val Val Met Ala Asn Tyr Glu Glu Ser Ile Lys Met Pro Ile Asn  
 225 230 235 240  
 Glu Pro Ala Pro Gly Lys Lys Lys Ser Gln Ile Gln Glu Tyr Val Asp  
 245 250 255  
 Tyr Asn Gly Gly Ala Gly Val Gln His Ile Ala Leu Lys Thr Glu Asp  
 260 265 270  
 Ile Ile Thr Ala Ile Arg Ser Leu Arg Glu Arg Gly Val Glu Phe Leu  
 275 280 285  
 Ala Val Pro Phe Thr Tyr Tyr Lys Gln Leu Gln Glu Lys Leu Lys Ser  
 290 295 300  
 Ala Lys Ile Arg Val Lys Glu Ser Ile Asp Val Leu Glu Glu Leu Lys  
 305 310 315 320  
 Ile Leu Val Asp Tyr Asp Glu Lys Gly Tyr Leu Leu Gln Ile Phe Thr  
 325 330 335  
 Lys Pro Met Gln Asp Arg Pro Thr Val Phe Leu Glu Val Ile Gln Arg  
 340 345 350  
 Asn Asn His Gln Gly Phe Gly Ala Gly Asn Phe Asn Ser Leu Phe Lys  
 355 360 365  
 Ala Phe Glu Glu Glu Gln Glu Leu Arg Gly Asn Leu Thr Asp Thr Asp  
 370 375 380  
 Pro Asn Gly Val Pro Phe Arg Leu  
 385 390

## (2) INFORMATION FOR SEQ ID NO:8:

- (i) SEQUENCE CHARACTERISTICS:
- (A) LENGTH: 392 amino acids
  - (B) TYPE: amino acid

(C) STRANDEDNESS: single  
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:8:

```

Thr Thr Tyr Asn Asn Lys Gly Pro Lys Pro Glu Arg Gly Arg Phe Leu
1      5      10      15
His Phe His Ser Val Thr Phe Trp Val Gly Asn Ala Lys Gln Ala Ala
20      25      30
Ser Phe Tyr Cys Asn Lys Met Gly Phe Glu Pro Leu Ala Tyr Arg Gly
35      40      45
Leu Glu Thr Gly Ser Arg Glu Val Val Ser His Val Ile Lys Arg Gly
50      55      60
Lys Ile Val Phe Val Leu Cys Ser Ala Leu Asn Pro Trp Asn Lys Glu
65      70      75      80
Met Gly Asp His Leu Val Lys His Gly Asp Gly Val Lys Asp Ile Ala
85      90      95
Phe Glu Val Glu Asp Cys Asp His Ile Val Gln Lys Ala Arg Glu Arg
100     105     110
Gly Ala Lys Ile Val Arg Glu Pro Trp Val Glu Gln Asp Lys Phe Gly
115     120     125
Lys Val Lys Phe Ala Val Leu Gln Thr Tyr Gly Asp Thr Thr His Thr
130     135     140
Leu Val Glu Lys Ile Asn Tyr Thr Gly Arg Phe Leu Pro Gly Phe Glu
145     150     155     160
Ala Pro Thr Tyr Lys Asp Thr Leu Leu Pro Lys Leu Pro Arg Cys Asn
165     170     175
Leu Glu Ile Ile Asp His Ile Val Gly Asn Gln Pro Asp Gln Glu Met
180     185     190
Gln Ser Ala Ser Glu Trp Tyr Leu Lys Asn Leu Gln Phe His Arg Phe
195     200     205
Trp Ser Val Asp Asp Thr Gln Val His Thr Glu Tyr Ser Ser Leu Arg
210     215     220
Ser Ile Val Val Thr Asn Tyr Glu Glu Ser Ile Lys Met Pro Ile Asn
225     230     235     240
Glu Pro Ala Pro Gly Arg Lys Lys Ser Gln Ile Gln Glu Tyr Val Asp
245     250     255
Tyr Asn Gly Gly Ala Gly Val Gln His Ile Ala Leu Lys Thr Glu Asp
260     265     270
Ile Ile Thr Ala Ile Arg His Leu Arg Glu Arg Gly Thr Glu Phe Leu
275     280     285
Ala Ala Pro Ser Ser Tyr Tyr Lys Leu Leu Arg Glu Asn Leu Lys Ser
290     295     300
Ala Lys Ile Gln Val Lys Glu Ser Met Asp Val Leu Glu Glu Leu His
305     310     315     320

```

(2) INFORMATION FOR SEQ ID NO:9:

```
(A) LENGTH: 376 amino acids
(B) TYPE: amino acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear
```

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:9:

44

Val Asp Asp Thr Gln Val His Thr Glu Tyr Ser Ser Leu Arg Ser Ile  
 210 215 220

Val Val Ala Asn Tyr Glu Glu Ser Ile Lys Met Pro Ile Asn Glu Pro  
 225 230 235 240

Ala Pro Gly Arg Lys Lys Ser Gln Ile Gln Glu Tyr Val Asp Tyr Asn  
 245 250 255

Gly Gly Ala Gly Val Gln His Ile Ala Leu Arg Thr Glu Asp Ile Ile  
 260 265 270

Thr Thr Ile Arg His Leu Arg Glu Arg Gly Met Glu Phe Leu Ala Val  
 275 280 285

Pro Ser Ser Tyr Tyr Arg Leu Leu Arg Glu Asn Leu Lys Thr Ser Lys  
 290 295 300

Ile Gln Val Lys Glu Asn Met Asp Val Leu Glu Glu Leu Lys Ile Leu  
 305 310 315 320

Val Asp Tyr Asp Glu Lys Gly Tyr Leu Leu Gln Ile Phe Thr Lys Pro  
 325 330 335

Met Gln Asp Arg Pro Thr Leu Phe Leu Glu Val Ile Gln Arg His Asn  
 340 345 350

His Gln Gly Phe Gly Ala Gly Asn Phe Asn Ser Leu Phe Lys Ala Phe  
 355 360 365

Glu Glu Glu Gln Ala Leu Arg Gly  
 370 375

## (2) INFORMATION FOR SEQ ID NO:10:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 1766 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: cDNA to mRNA
- (iii) HYPOTHETICAL: NO
- (iv) ANTI-SENSE: NO
- (vi) ORIGINAL SOURCE:
  - (A) ORGANISM: Zea mays
- (ix) FEATURE:
  - (A) NAME/KEY: CDS
  - (B) LOCATION: 261..1595
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:10:

ACTAGTTGTG AGAGCCTTCT GCGTTGGCAA TTGGCAGTAC AAGACAAATC ACATCCGCAA 60

CCGCAACCAC AGAATCGTCC GTCCACGTGG CCCCCATCAC TTCCCTTTAT TTACCAGTCG 120

TCCCCCATCC CCAGGGCCAC CCACCAACAA GTGCAGTCAC CCGAGCCGCA AACTGCAGCT 180

CTGCAAGCTA CAGAGGCCAC CACGAGTCCA CGACGCCACG CCCTCCGAGA GAAAGAGAAA 240

GAGAAAACCA AAGCAGGATA ATG CCC CCG ACC CCC ACA GCC GCC GCA GCC	290
Met Pro Pro Thr Pro Thr Ala Ala Ala Ala	
1 5 10	
GGC GCC GCC GTG GCG GCG GCA TCA GCA GCG GAG CAA GCG GCG TTC CGC	338
Gly Ala Ala Val Ala Ala Ala Ser Ala Ala Glu Gln Ala Ala Phe Arg	
15 20 25	
CTC GTG GGC CAC GCG AAC TTC GTC CCG TTC AAC CCG GCG TCC GAC CGC	386
Leu Val Gly His Arg Asn Phe Val Arg Phe Asn Pro Arg Ser Asp Arg	
30 35 40	
TTC CAC ACG CTC GCG TTC CAC CAC GTG GAG CTC TGG TGC GCC GAC GCG	434
Phe His Thr Leu Ala Phe His His Val Glu Leu Trp Cys Ala Asp Ala	
45 50 55	
GCC TCC GCC GCG GGC CCG TTC TCC TTC GGC CTG GGC GCG CCG CTC GCC	482
Ala Ser Ala Ala Gly Arg Phe Ser Phe Gly Leu Gly Ala Pro Leu Ala	
60 65 70	
GCA CGC TCC GAC CTC TCC ACG GGC AAC TCC GCG CAC GCG TCC CTG CTG	530
Ala Arg Ser Asp Leu Thr Gly Asn Ser Ala His Ala Ser Leu Leu	
75 80 85 90	
CTC CGC TCC GGC TCC CTC TCC TTC CTC TTC ACG GCG CCC TAC GCG CAC	578
Leu Arg Ser Gly Ser Leu Ser Phe Leu Thr Ala Pro Tyr Ala His	
95 100 105	
GGC GCC GAC GCT GCC ACC GCC GCG CTG CCC TCC TTC TCC GCC GCC GCC	626
Gly Ala Asp Ala Ala Thr Ala Ala Leu Pro Ser Phe Ser Ala Ala Ala	
110 115 120	
GCG CGG CGC TTC GCA GCC GAC CAC GGC CTC GCG GTG CGC GCC GTC GCG	674
Ala Arg Arg Phe Ala Ala Asp His Gly Leu Ala Val Arg Ala Val Ala	
125 130 135	
CTC CGC GTC GCC GAC GCC GAG GAC GCC TTC CGC GCC AGC GTC GCG GCC	722
Leu Arg Val Ala Asp Ala Glu Asp Ala Phe Arg Ala Ser Val Ala Ala	
140 145 150	
GGG GCG CGC CCG GCG TTC GGC CCC GTC GAC CTC GGC CGC GGC TTC CGC	770
Gly Ala Arg Pro Ala Phe Gly Pro Val Asp Leu Gly Arg Gly Phe Arg	
155 160 165 170	
CTC GCC GAG GTC GAG CTC TAC GGC GAC GTC GTG CTC CGG TAC GTG AGC	818
Leu Ala Glu Val Glu Leu Tyr Gly Asp Val Val Leu Arg Tyr Val Ser	
175 180 185	
TAC CCG GAC GGC GCC GCG GGC GAG CCC TTC CTG CCG GGG TTC GAG GGC	866
Tyr Pro Asp Gly Ala Ala Gly Glu Pro Phe Leu Pro Gly Phe Glu Gly	
190 195 200	
GTG GCC AGC CCC GGG GCG GCC GAC TAC GGG CTG AGC AGG TTC GAC CAC	914
Val Ala Ser Pro Gly Ala Ala Asp Tyr Gly Leu Ser Arg Phe Asp His	
205 210 215	
ATC GTC GGC AAC GTG CCG GAG CTG GCG CCC GCC GCC GCC TAC TTC GCC	962
Ile Val Gly Asn Val Pro Glu Leu Ala Pro Ala Ala Ala Tyr Phe Ala	
220 225 230	
GGC TTC ACG GGG TTC CAC GAG TTC GCC GAG TTC ACG ACG GAG GAC GTG	1010
Gly Phe Thr Gly Phe His Glu Phe Ala Glu Phe Thr Thr Glu Asp Val	
235 240 245 250	



GGC ACC GCG GAG AGC GGC CTC AAC TCC ATG GTG CTC GCC AAC AAC TCG	1058
Gly Thr Ala Glu Ser Gly Leu Asn Ser Met Val Leu Ala Asn Asn Ser	
255 260 265	
GAG AAC GTG CTG CTC CCG CTC AAC GAG CCG GTG CAC GGC ACC AAG CGC	1106
Glu Asn Val Leu Leu Pro Leu Asn Glu Pro Val His Gly Thr Lys Arg	
270 275 280	
CGC AGC CAG ATA CAA ACG TTC CTG GAC CAC CAC GGC GGC CCC GGC GTG	1154
Arg Ser Gln Ile Gln Thr Phe Leu Asp His His Gly Gly Pro Gly Val	
285 290 295	
CAG CAC ATG GCG CTG GCC AGC GAC GAC GTG CTC AGG ACG CTG AGG GAG	1202
Gln His Met Ala Leu Ala Ser Asp Asp Val Leu Arg Thr Leu Arg Glu	
300 305 310	
ATG CAG GCG CGC TCG GCC ATG GGC GGC TTC GAG TTC ATG GCG CCT CCC	1250
Met Gln Ala Arg Ser Ala Met Gly Gly Phe Glu Phe Met Ala Pro Pro	
315 320 325 330	
ACA TCC GAC TAC TAT GAC GGC GTG AGG CGG CGC GCC GGG GAC GTG CTC	1298
Thr Ser Asp Tyr Tyr Asp Gly Val Arg Arg Arg Ala Gly Asp Val Leu	
335 340 345	
ACG GAA GCA CAG ATT AAG GAG TGC CAG GAG CTA GGG GTG CTG GTG GAC	1346
Thr Glu Ala Gln Ile Lys Glu Cys Gln Glu Leu Gly Val Leu Val Asp	
350 355 360	
AGG GAT GAC CAG GGC GTG CTG CTC CAA ATC TTC ACC AAG CCA GTG GGG	1394
Arg Asp Asp Gln Gly Val Leu Gln Ile Phe Thr Lys Pro Val Gly	
365 370 375	
GAC AGG CCA ACG CTG TTC TTG GAA ATC ATC CAA AGG ATC GGG TGC ATG	1442
Asp Arg Pro Thr Leu Phe Leu Glu Ile Ile Gln Arg Ile Gly Cys Met	
380 385 390	
GAG AAG GAT GAG AAG GGG CAA GAA TAC CAA AAG GGT GGC TGC GGC GGG	1490
Glu Lys Asp Glu Lys Gly Gln Glu Tyr Gln Lys Gly Gly Cys Gly Gly	
395 400 405 410	
TTC GGC AAG GGA AAC TTC TCG CAG CTG TTC AAG TCC ATC GAG GAT TAT	1538
Phe Gly Lys Gly Asn Phe Ser Gln Leu Phe Lys Ser Ile Glu Asp Tyr	
415 420 425	
GAG AAG TCC CTT GAA GCC AAG CAA GCT GCT GCA GCA GCT GCA GCT CAG	1586
Glu Lys Ser Leu Glu Ala Lys Gln Ala Ala Ala Ala Ala Ala Gln	
430 435 440	
GGA TCC TAG GACAGTGCTT GGAGACGAGC AACTGCTGTG GCACTTTGTA	1635
Gly Ser	
TCATGGAACA GAAATAATGA AGCGTGTCT TTGTGACACT TGACATGCAA ATGTTTGTGT	1695
TCTGTAACCG TTGAATATAT GGGACGATGC TATGATGGTG TAATAGATGG TAGAGAGGGT	1755
ACAACCCTGA T	1766

## (2) INFORMATION FOR SEQ ID NO:11:

## (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 445 amino acids
- (B) TYPE: amino acid
- (D) TOPOLOGY: linear

## (ii) MOLECULE TYPE: protein

## (xi) SEQUENCE DESCRIPTION: SEQ ID NO:11:

```

Met Pro Pro Thr Pro Thr Ala Ala Ala Ala Gly Ala Ala Val Ala Ala
 1           5           10           15

Ala Ser Ala Ala Glu Gln Ala Ala Phe Arg Leu Val Gly His Arg Asn
          20           25           30

Phe Val Arg Phe Asn Pro Arg Ser Asp Arg Phe His Thr Leu Ala Phe
          35           40           45

His His Val Glu Leu Trp Cys Ala Asp Ala Ala Ser Ala Ala Gly Arg
 50           55           60

Phe Ser Phe Gly Leu Gly Ala Pro Leu Ala Ala Arg Ser Asp Leu Ser
 65           70           75           80

Thr Gly Asn Ser Ala His Ala Ser Leu Leu Leu Arg Ser Gly Ser Leu
          85           90           95

Ser Phe Leu Phe Thr Ala Pro Tyr Ala His Gly Ala Asp Ala Ala Thr
          100          105          110

Ala Ala Leu Pro Ser Phe Ser Ala Ala Ala Arg Arg Phe Ala Ala
          115          120          125

Asp His Gly Leu Ala Val Arg Ala Val Ala Leu Arg Val Ala Asp Ala
          130          135          140

Glu Asp Ala Phe Arg Ala Ser Val Ala Ala Gly Ala Arg Pro Ala Phe
          145          150          155          160

Gly Pro Val Asp Leu Gly Arg Gly Phe Arg Leu Ala Glu Val Glu Leu
          165          170          175

Tyr Gly Asp Val Val Leu Arg Tyr Val Ser Tyr Pro Asp Gly Ala Ala
          180          185          190

Gly Glu Pro Phe Leu Pro Gly Phe Glu Gly Val Ala Ser Pro Gly Ala
          195          200          205

Ala Asp Tyr Gly Leu Ser Arg Phe Asp His Ile Val Gly Asn Val Pro
          210          215          220

Glu Leu Ala Pro Ala Ala Ala Tyr Phe Ala Gly Phe Thr Gly Phe His
          225          230          235          240

Glu Phe Ala Glu Phe Thr Thr Glu Asp Val Gly Thr Ala Glu Ser Gly
          245          250          255

Leu Asn Ser Met Val Leu Ala Asn Asn Ser Glu Asn Val Leu Leu Pro
          260          265          270

Leu Asn Glu Pro Val His Gly Thr Lys Arg Arg Ser Gln Ile Gln Thr
          275          280          285

Phe Leu Asp His His Gly Gly Pro Gly Val Gln His Met Ala Leu Ala
          290          295          300

Ser Asp Asp Val Leu Arg Thr Leu Arg Glu Met Gln Ala Arg Ser Ala
          305          310          315          320

Met Gly Gly Phe Glu Phe Met Ala Pro Pro Thr Ser Asp Tyr Tyr Asp
          325          330          335

```

Gly Val Arg Arg Arg Ala Gly Asp Val Leu Thr Glu Ala Gln Ile Lys  
 340 345 350  
 Glu Cys Gln Glu Leu Gly Val Leu Val Asp Arg Asp Asp Gln Gly Val  
 355 360 365  
 Leu Leu Gln Ile Phe Thr Lys Pro Val Gly Asp Arg Pro Thr Leu Phe  
 370 375 380  
 Leu Glu Ile Ile Gln Arg Ile Gly Cys Met Glu Lys Asp Glu Lys Gly  
 385 390 395 400  
 Gln Glu Tyr Gln Lys Gly Gly Cys Gly Gly Phe Gly Lys Gly Asn Phe  
 405 410 415  
 Ser Gln Leu Phe Lys Ser Ile Glu Asp Tyr Glu Lys Ser Leu Glu Ala  
 420 425 430  
 Lys Gln Ala Ala Ala Ala Ala Ala Ala Gln Gly Ser  
 435 440

## (2) INFORMATION FOR SEQ ID NO:12:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 1356 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: double
  - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: cDNA to mRNA
- (iii) HYPOTHETICAL: NO
- (vi) ORIGINAL SOURCE:
  - (A) ORGANISM: Arabidopsis thaliana
- (ix) FEATURE:
  - (A) NAME/KEY: CDS
  - (B) LOCATION: 1..1254
- (ix) FEATURE:
  - (A) NAME/KEY: misc\_feature
  - (B) LOCATION: 1..3
  - (D) OTHER INFORMATION: /standard\_name="translation initiation codon"
- (ix) FEATURE:
  - (A) NAME/KEY: misc\_feature
  - (B) LOCATION: 1252..1254
  - (D) OTHER INFORMATION: /standard\_name="translation termination codon"

## (xi) SEQUENCE DESCRIPTION: SEQ ID NO:12:

ATG TCC AAG TTC GTA AGA AAG AAT CCA AAG TCT GAT AAA TTC AAG GTT	48
Met Ser Lys Phe Val Arg Lys Asn Pro Lys Ser Asp Lys Phe Lys Val	
1 5 10 15	
AAG CGC TTC CAT CAC ATC GAG TTC TGG TGC GGC GAC GCA ACC AAC GTC	96
Lys Arg Phe His His Ile Glu Phe Trp Cys Gly Asp Ala Thr Asn Val	
20 25 30	

GCT CGT CGC TTC TCC TGG GGT CTG GGG ATG AGA TTC TCC GCC AAA TCC	144
Ala Arg Arg Phe Ser Trp Gly Leu Gly Met Arg Phe Ser Ala Lys Ser	
35 40 45	
GAT CTT TCC ACC GGA AAC ATG GTT CAC GCC TCT TAC CTA CTC ACC TCC	192
Asp Leu Ser Thr Gly Asn Met Val His Ala Ser Tyr Leu Leu Thr Ser	
50 55 60	
GGT GAC CTC CGA TTC CTT TTC ACT GCT CCT TAC TCT CCG TCT CTC TCC	240
Gly Asp Leu Arg Phe Leu Phe Thr Ala Pro Tyr Ser Pro Ser Leu Ser	
65 70 75 80	
GCC GGA GAG ATT AAA CCG ACA ACC ACA GCT TCT ATC CCA AGT TTC GAT	288
Ala Gly Glu Ile Lys Pro Thr Thr Thr Ala Ser Ile Pro Ser Phe Asp	
85 90 95	
CAC GGC TCT TGT CGT TCC TTC TTC TCT TCA CAT GGT CTC GGT GTT AGA	336
His Gly Ser Cys Arg Ser Phe Phe Ser Ser His Gly Leu Gly Val Arg	
100 105 110	
GCC GTT GCG ATT GAA GTA GAA GAC GCA GAG TCA GCT TTC TCC ATC AGT	384
Ala Val Ala Ile Glu Val Glu Asp Ala Glu Ser Ala Phe Ser Ile Ser	
115 120 125	
GTA GCT AAT GGC GCT ATT CCT TCG TCG CCT CCT ATC GTC CTC AAT GAA	432
Val Ala Asn Gly Ala Ile Pro Ser Ser Pro Pro Ile Val Leu Asn Glu	
130 135 140	
GCA GTT ACG ATC GCT GAG GTT AAA CTA TAC GGC GAT GTT GTT CTC CGA	480
Ala Val Thr Ile Ala Glu Val Lys Leu Tyr Gly Asp Val Val Leu Arg	
145 150 155 160	
TAT GTT AGT TAC AAA GCA GAA GAT ACC GAA AAA TCC GAA TTC TTG CCA	528
Tyr Val Ser Tyr Lys Ala Glu Asp Thr Glu Lys Ser Glu Phe Leu Pro	
165 170 175	
GGG TTC GAG CGT GTA GAG GAT GCG TCG TCG TTC CCA TTG GAT TAT GGT	576
Gly Phe Glu Arg Val Glu Asp Ala Ser Ser Phe Pro Leu Asp Tyr Gly	
180 185 190	
ATC CGG CGG CTT GAC CAC GCC GTG GGA AAC GTT CCT GAG CTT GGT CCG	624
Ile Arg Arg Leu Asp His Ala Val Gly Asn Val Pro Glu Leu Gly Pro	
195 200 205	
GCT TTA ACT TAT GTA GCG GGG TTC ACT GGT TTT CAC CAA TTC GCA GAG	672
Ala Leu Thr Tyr Val Ala Gly Phe Thr Gly Phe His Gln Phe Ala Glu	
210 215 220	
TTC ACA GCA GAC GAC GTT GGA ACC GCC GAG AGC GGT TTA AAT TCA GCG	720
Phe Thr Ala Asp Asp Val Gly Thr Ala Glu Ser Gly Leu Asn Ser Ala	
225 230 235 240	
GTC CTG GCT AGC AAT GAT GAA ATG GTT CTT CTA CCG ATT AAC GAG CCA	768
Val Leu Ala Ser Asn Asp Glu Met Val Leu Leu Pro Ile Asn Glu Pro	
245 250 255	
GTG CAC GGA ACA AAG AGG AAG AGT CAG ATT CAG ACG TAT TTG GAA CAT	816
Val His Gly Thr Lys Arg Lys Ser Gln Ile Gln Thr Tyr Leu Glu His	
260 265 270	
AAC GAA GGC GCA GGG CTA CAA CAT CTG GCT CTG ATG AGT GAA GAC ATA	864
Asn Glu Gly Ala Gly Leu Gln His Leu Ala Leu Met Ser Glu Asp Ile	
275 280 285	

TTC AGG ACC CTG AGA GAG ATG AGG AAG AGG AGC AGT ATT GGA GGA TTC 912  
 Phe Arg Thr Leu Arg Glu Met Arg Lys Arg Ser Ser Ile Gly Gly Phe  
 290 295 300

GAC TTC ATG CCT TCT CCT CCG CCT ACT TAC TAC CAG AAT CTC AAG AAA 960  
 Asp Phe Met Pro Ser Pro Pro Pro Thr Tyr Tyr Gln Asn Leu Lys Lys  
 305 310 315 320

CGG GTC GGC GAC GTG CTC AGC GAT GAT CAG ATC AAG GAG TGT GAG GAA 1008  
 Arg Val Gly Asp Val Leu Ser Asp Asp Gln Ile Lys Glu Cys Glu Glu  
 325 330 335

TTA GGG ATT CTT GTA GAC AGA GAT GAT CAA GGG ACG TTG CTT CAA ATC 1056  
 Leu Gly Ile Leu Val Asp Arg Asp Asp Gln Gly Thr Leu Leu Gln Ile  
 340 345 350

TTC ACA AAA CCA CTA GGT GAC AGG CCG ACG ATA TTT ATA GAG ATA ATC 1104  
 Phe Thr Lys Pro Leu Gly Asp Arg Pro Thr Ile Phe Ile Glu Ile Ile  
 355 360 365

CAG AGA GTA GGA TGC ATG ATG AAA GAT GAG GAA GGG AAG GCT TAC CAG 1152  
 Gln Arg Val Gly Cys Met Met Lys Asp Glu Glu Gly Lys Ala Tyr Gln  
 370 375 380

AGT GGA GGA TGT GGT GGT TTT GGC AAA GGC AAT TTC TCT GAG CTC TTC 1200  
 Ser Gly Gly Cys Gly Phe Gly Lys Gly Asn Phe Ser Glu Leu Phe  
 385 390 395 400

AAG TCC ATT GAA GAA TAC GAA AAG ACT CTT GAA GCC AAA CAG TTA GTG 1248  
 Lys Ser Ile Glu Glu Tyr Glu Lys Thr Leu Glu Ala Lys Gln Leu Val  
 405 410 415

GGA TGA ACAAGAAGAA GAACCAACTA AAGGATTGTG TAATTAATGT AAAACTGTTT 1304  
 Gly \*

TATCTTATCA AAACAATGTA TACAACATCT CATTAAAAA CGAGATCAAT CC 1356

## (2) INFORMATION FOR SEQ ID NO:13:

## (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 418 amino acids  
 (B) TYPE: amino acid  
 (D) TOPOLOGY: linear

## (ii) MOLECULE TYPE: protein

## (xi) SEQUENCE DESCRIPTION: SEQ ID NO:13:

Met Ser Lys Phe Val Arg Lys Asn Pro Lys Ser Asp Lys Phe Lys Val  
 1 5 10 15

Lys Arg Phe His His Ile Glu Phe Trp Cys Gly Asp Ala Thr Asn Val  
 20 25 30

Ala Arg Arg Phe Ser Trp Gly Leu Gly Met Arg Phe Ser Ala Lys Ser  
 35 40 45

Asp Leu Ser Thr Gly Asn Met Val His Ala Ser Tyr Leu Leu Thr Ser  
 50 55 60

Gly Asp Leu Arg Phe Leu Phe Thr Ala Pro Tyr Ser Pro Ser Leu Ser  
 65 70 75 80

Ala Gly Glu Ile Lys Pro Thr Thr Thr Ala Ser Ile Pro Ser Phe Asp  
 85 90 95

His Gly Ser Cys Arg Ser Phe Phe Ser Ser His Gly Leu Gly Val Arg  
 100 105 110  
 Ala Val Ala Ile Glu Val Glu Asp Ala Glu Ser Ala Phe Ser Ile Ser  
 115 120 125  
 Val Ala Asn Gly Ala Ile Pro Ser Ser Pro Pro Ile Val Leu Asn Glu  
 130 135 140  
 Ala Val Thr Ile Ala Glu Val Lys Leu Tyr Gly Asp Val Val Leu Arg  
 145 150 155 160  
 Tyr Val Ser Tyr Lys Ala Glu Asp Thr Glu Lys Ser Glu Phe Leu Pro  
 165 170 175  
 Gly Phe Glu Arg Val Glu Asp Ala Ser Ser Phe Pro Leu Asp Tyr Gly  
 180 185 190  
 Ile Arg Arg Leu Asp His Ala Val Gly Asn Val Pro Glu Leu Gly Pro  
 195 200 205  
 Ala Leu Thr Tyr Val Ala Gly Phe Thr Gly Phe His Gln Phe Ala Glu  
 210 215 220  
 Phe Thr Ala Asp Asp Val Gly Thr Ala Glu Ser Gly Leu Asn Ser Ala  
 225 230 235 240  
 Val Leu Ala Ser Asn Asp Glu Met Val Leu Leu Pro Ile Asn Glu Pro  
 245 250 255  
 Val His Gly Thr Lys Arg Lys Ser Gln Ile Gln Thr Tyr Leu Glu His  
 260 265 270  
 Asn Glu Gly Ala Gly Leu Gln His Leu Ala Leu Met Ser Glu Asp Ile  
 275 280 285  
 Phe Arg Thr Leu Arg Glu Met Arg Lys Arg Ser Ser Ile Gly Gly Phe  
 290 295 300  
 Asp Phe Met Pro Ser Pro Pro Pro Thr Tyr Tyr Gln Asn Leu Lys Lys  
 305 310 315 320  
 Arg Val Gly Asp Val Leu Ser Asp Asp Gln Ile Lys Glu Cys Glu Glu  
 325 330 335  
 Leu Gly Ile Leu Val Asp Arg Asp Asp Gln Gly Thr Leu Leu Gln Ile  
 340 345 350  
 Phe Thr Lys Pro Leu Gly Asp Arg Pro Thr Ile Phe Ile Glu Ile Ile  
 355 360 365  
 Gln Arg Val Gly Cys Met Met Lys Asp Glu Glu Gly Lys Ala Tyr Gln  
 370 375 380  
 Ser Gly Gly Cys Gly Gly Phe Gly Lys Gly Asn Phe Ser Glu Leu Phe  
 385 390 395 400  
 Lys Ser Ile Glu Glu Tyr Glu Lys Thr Leu Glu Ala Lys Gln Leu Val  
 405 410 415  
 Gly \*

## (2) INFORMATION FOR SEQ ID NO:14:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 1448 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: double
  - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: cDNA to mRNA
- (iii) HYPOTHETICAL: NO
- (vi) ORIGINAL SOURCE:
  - (A) ORGANISM: Arabidopsis thaliana
- (ix) FEATURE:
  - (A) NAME/KEY: CDS
  - (B) LOCATION: 9..1346
- (ix) FEATURE:
  - (A) NAME/KEY: misc feature
  - (B) LOCATION: 9..11
  - (D) OTHER INFORMATION: /standard\_name="translation initiation codon"
- (ix) FEATURE:
  - (A) NAME/KEY: misc feature
  - (B) LOCATION: 1344..1346
  - (D) OTHER INFORMATION: /standard\_name="translation termination codon"

## (xi) SEQUENCE DESCRIPTION: SEQ ID NO:14:

```

TGAAATCA ATG GGC CAC CAA AAC GCC GCC GTT TCA GAG AAT CAA AAC CAT      50
      Met Gly His Gln Asn Ala Ala Val Ser Glu Asn Gln Asn His
        1              5              10

GAT GAC GGC GCT GCG TCG TCG CCG GGA TTC AAG CTC GTC GGA TTT TCC      98
Asp Asp Gly Ala Ala Ser Ser Pro Gly Phe Lys Leu Val Gly Phe Ser
  15              20              25              30

AAG TTC GTA AGA AAG AAT CCA AAG TCT GAT AAA TTC AAG GTT AAG CGC      146
Lys Phe Val Arg Lys Asn Pro Lys Ser Asp Lys Phe Lys Val Lys Arg
          35              40              45

TTC CAT CAC ATC GAG TTC TGG TGC GGC GAC GCA ACC AAC GTC GCT CGT      194
Phe His His Ile Glu Phe Trp Cys Gly Asp Ala Thr Asn Val Ala Arg
          50              55              60

CGC TTC TCC TGG GGT CTG GGG ATG AGA TTC TCC GCC AAA TCC GAT CTT      242
Arg Phe Ser Trp Gly Leu Gly Met Arg Phe Ser Ala Lys Ser Asp Leu
        65              70              75

TCC ACC GGA AAC ATG GTT CAC GCC TCT TAC CTA CTC ACC TCC GGT GAC      290
Ser Thr Gly Asn Met Val His Ala Ser Tyr Leu Leu Thr Ser Gly Asp
        80              85              90

CTC CGA TTC CTT TTC ACT GCT CCT TAC TCT CCG TCT CTC TCC GCC GGA      338
Leu Arg Phe Leu Phe Thr Ala Pro Tyr Ser Pro Ser Leu Ser Ala Gly
        95              100              105              110

GAG ATT AAA CCG ACA ACC ACA GCT TCT ATC CCA AGT TTC GAT CAC GGC      386
Glu Ile Lys Pro Thr Thr Thr Ala Ser Ile Pro Ser Phe Asp His Gly
          115              120              125

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TCT TGT CGT TCC TTC TTC TCT TCA CAT GGT CTC GGT GTT AGA GCC GTT	434
Ser Cys Arg Ser Phe Phe Ser Ser His Gly Leu Gly Val Arg Ala Val	
130 135 140	
GCG ATT GAA GTA GAA GAC GCA GAG TCA GCT TTC TCC ATC AGT GTA GCT	482
Ala Ile Glu Val Glu Asp Ala Glu Ser Ala Phe Ser Ile Ser Val Ala	
145 150 155	
AAT GGC GCT ATT CCT TCG TCG CCT CCT ATC GTC CTC AAT GAA GCA GTT	530
Asn Gly Ala Ile Pro Ser Ser Pro Pro Ile Val Leu Asn Glu Ala Val	
160 165 170	
ACG ATC GCT GAG GTT AAA CTA TAC GGC GAT GTT GTT CTC CGA TAT GTT	578
Thr Ile Ala Glu Val Lys Leu Tyr Gly Asp Val Val Leu Arg Tyr Val	
175 180 185 190	
AGT TAC AAA GCA GAA GAT ACC GAA AAA TCC GAA TTC TTG CCA GGG TTC	626
Ser Tyr Lys Ala Glu Asp Thr Glu Lys Ser Glu Phe Leu Pro Gly Phe	
195 200 205	
GAG CGT GTA GAG GAT GCG TCG TCG TTC CCA TTG GAT TAT GGT ATC CGG	674
Glu Arg Val Glu Asp Ala Ser Ser Phe Pro Leu Asp Tyr Gly Ile Arg	
210 215 220	
CGG CTT GAC CAC GCC GTG GGA AAC GTT CCT GAG CTT GGT CCG GCT TTA	722
Arg Leu Asp His Ala Val Gly Asn Val Pro Glu Leu Gly Pro Ala Leu	
225 230 235	
ACT TAT GTA GCG GGG TTC ACT GGT TTT CAC CAA TTC GCA GAG TTC ACA	770
Thr Tyr Val Ala Gly Phe Thr Gly Phe His Gln Phe Ala Glu Phe Thr	
240 245 250	
GCA GAC GAC GTT GGA ACC GCC GAG AGC GGT TTA AAT TCA GCG GTC CTG	818
Ala Asp Asp Val Gly Thr Ala Glu Ser Gly Leu Asn Ser Ala Val Leu	
255 260 265 270	
GCT AGC AAT GAT GAA ATG GTT CTT CTA CCG ATT AAC GAG CCA GTG CAC	866
Ala Ser Asn Asp Glu Met Val Leu Leu Pro Ile Asn Glu Pro Val His	
275 280 285	
GGA ACA AAG AGG AAG AGT CAG ATT CAG ACG TAT TTG GAA CAT AAC GAA	914
Gly Thr Lys Arg Lys Ser Gln Ile Gln Thr Tyr Leu Glu His Asn Glu	
290 295 300	
GGC GCA GGG CTA CAA CAT CTG GCT CTG ATG AGT GAA GAC ATA TTC AGG	962
Gly Ala Gly Leu Gln His Leu Ala Leu Met Ser Glu Asp Ile Phe Arg	
305 310 315	
ACC CTG AGA GAG ATG AGG AAG AGG AGC AGT ATT GGA GGA TTC GAC TTC	1010
Thr Leu Arg Glu Met Arg Lys Arg Ser Ser Ile Gly Gly Phe Asp Phe	
320 325 330	
ATG CCT TCT CCT CCG CCT ACT TAC TAC CAG AAT CTC AAG AAA CGG GTC	1058
Met Pro Ser Pro Pro Pro Thr Tyr Tyr Gln Asn Leu Lys Lys Arg Val	
335 340 345 350	
GGC GAC GTG CTC AGC GAT GAT CAG ATC AAG GAG TGT GAG GAA TTA GGG	1106
Gly Asp Val Leu Ser Asp Asp Gln Ile Lys Glu Cys Glu Glu Leu Gly	
355 360 365	
ATT CTT GTA GAC AGA GAT GAT CAA GGG ACG TTG CTT CAA ATC TTC ACA	1154
Ile Leu Val Asp Arg Asp Asp Gln Gly Thr Leu Leu Gln Ile Phe Thr	
370 375 380	



AAACAATGTA TACAACATCT CATTTAAAAA CGAGATCAAT CC 1448

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:15:

Ala Glu Val Lys Leu Tyr Gly Asp Val Val Leu Arg Tyr Val Ser Tyr  
180 185 190

Lys Ala Glu Asp Thr Glu Lys Ser Glu Phe Leu Pro Gly Phe Glu Arg  
 195 200 205  
 Val Glu Asp Ala Ser Ser Phe Pro Leu Asp Tyr Gly Ile Arg Arg Leu  
 210 215 220  
 Asp His Ala Val Gly Asn Val Pro Glu Leu Gly Pro Ala Leu Thr Tyr  
 225 230 235 240  
 Val Ala Gly Phe Thr Gly Phe His Gln Phe Ala Glu Phe Thr Ala Asp  
 245 250 255  
 Asp Val Gly Thr Ala Glu Ser Gly Leu Asn Ser Ala Val Leu Ala Ser  
 260 265 270  
 Asn Asp Glu Met Val Leu Leu Pro Ile Asn Glu Pro Val His Gly Thr  
 275 280 285  
 Lys Arg Lys Ser Gln Ile Gln Thr Tyr Leu Glu His Asn Glu Gly Ala  
 290 295 300  
 Gly Leu Gln His Leu Ala Leu Met Ser Glu Asp Ile Phe Arg Thr Leu  
 305 310 315 320  
 Arg Glu Met Arg Lys Arg Ser Ser Ile Gly Gly Phe Asp Phe Met Pro  
 325 330 335  
 Ser Pro Pro Pro Thr Tyr Tyr Gln Asn Leu Lys Lys Arg Val Gly Asp  
 340 345 350  
 Val Leu Ser Asp Asp Gln Ile Lys Glu Cys Glu Glu Leu Gly Ile Leu  
 355 360 365  
 Val Asp Arg Asp Asp Gln Gly Thr Leu Leu Gln Ile Phe Thr Lys Pro  
 370 375 380  
 Leu Gly Asp Arg Pro Thr Ile Phe Ile Glu Ile Ile Gln Arg Val Gly  
 385 390 395 400  
 Cys Met Met Lys Asp Glu Glu Gly Lys Ala Tyr Gln Ser Gly Gly Cys  
 405 410 415  
 Gly Gly Phe Gly Lys Gly Asn Phe Ser Glu Leu Phe Lys Ser Ile Glu  
 420 425 430  
 Glu Tyr Glu Lys Thr Leu Glu Ala Lys Gln Leu Val Gly \*  
 435 440 445

## (2) INFORMATION FOR SEQ ID NO:16:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 513 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: double
  - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: cDNA to mRNA
- (iii) HYPOTHETICAL: NO
- (vi) ORIGINAL SOURCE:
  - (A) ORGANISM: Vernonia galamenensis
- (vii) IMMEDIATE SOURCE:
  - (B) CLONE: vs1.pk0015.b2

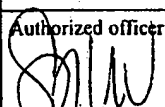
## (xi) SEQUENCE DESCRIPTION: SEQ ID NO:16:

CCACACCGAT TGCCGGAAC TCACCGCCTC TCACGGCCTT GCAGTCCGAG CAATCGCCAT	60
TGAAGTCGAT GACGCCGAAT TAGCTTTCTC CGTCAGCGTC TCTCACGGCG CTAAACCCCTC	120
CGCTGCTCCT GTAACCCCTTG GAAACAACGA CGTCGTATTG TCTGAAGTTA AGCTTTACGG	180
CGATGTCGCT TTCCGGTACA TAAGTTACAA AAATCCGAAC TATACATCTT CCTTTTTGCC	240
CGGGTTCGAG CCCGTTGAAA AGACGTCGTC GTTTTATGAC CTTGACTACG GTATCCGCCG	300
TTTGGACCAC GCCGTAGGNA ACGTCCCTGA GCTTGCTTCG GCAGTGGACT ACGTGAAATC	360
ATTCACCGGA TTCCATGAGT TCGCCGAATT CACCGCGGAG GACGTCGGGA CGAGCGAGAG	420
GGAAGTGAAT TCGGTCGTTT TAGCTTGCAA CAGTGAGATG GTCTTGATTC CGATGAACGA	480
GCCGGTGTAC GGAANAAAAG GAAGNAGCCA GAT	513

## INDICATIONS RELATING TO A DEPOSITED MICROORGANISM

(PCT Rule 13bis)

A. The indications made below relate to the microorganism referred to in the description on page <u>6</u> , line <u>1</u>	
B. IDENTIFICATION OF DEPOSIT <span style="float: right;">Further deposits are identified on an additional sheet <input type="checkbox"/></span>	
Name of depositary institution AMERICAN TYPE CULTURE COLLECTION	
Address of depositary institution (including postal code and country) 12301 Parklawn Drive Rockville, Maryland 20852 US	
Date of deposit 25 June 1996 (25.06.96)	Accession Number 98083
C. ADDITIONAL INDICATIONS (leave blank if not applicable) <span style="float: right;">This information is continued on an additional sheet <input checked="" type="checkbox"/></span>	
In respect of those designations in which a European patent is sought, a sample of the deposited microorganism will be made available until the publication of the mention of the grant of the European patent or until the date on which the application has been refused or withdrawn or is deemed to be withdrawn, only by the issue of such a sample to an expert nominated by the person requesting the sample. (Rule 28(4) EPC)	
D. DESIGNATED STATES FOR WHICH INDICATIONS ARE MADE (if the indications are not for all designated States)	
E. SEPARATE FURNISHING OF INDICATIONS (leave blank if not applicable)	
The indications listed below will be submitted to the International Bureau later (specify the general nature of the indications e.g. "Accession Number of Deposit")	

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<input checked="" type="checkbox"/> This sheet was received with the international application
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Authorized officer

## INDICATIONS RELATING TO A DEPOSITED MICROORGANISM

(PCT Rule 13bis)

<b>A.</b> The indications made below relate to the microorganism referred to in the description on page <u>6</u> , line <u>1</u>	
<b>B. IDENTIFICATION OF DEPOSIT</b> <span style="float: right;">Further deposits are identified on an additional sheet <input type="checkbox"/></span>	
Name of depositary institution <b>AMERICAN TYPE CULTURE COLLECTION</b>	
Address of depositary institution (including postal code and country) 12301 Parklawn Drive Rockville, Maryland 20852 US	
Date of deposit 25 June 1996 (25.06.96)	Accession Number 97622
<b>C. ADDITIONAL INDICATIONS</b> (leave blank if not applicable) <span style="float: right;">This information is continued on an additional sheet <input checked="" type="checkbox"/></span>	
In respect of those designations in which a European patent is sought, a sample of the deposited microorganism will be made available until the publication of the mention of the grant of the European patent or until the date on which the application has been refused or withdrawn or is deemed to be withdrawn, only by the issue of such a sample to an expert nominated by the person requesting the sample. (Rule 28(4) EPC)	
<b>D. DESIGNATED STATES FOR WHICH INDICATIONS ARE MADE</b> (if the indications are not for all designated States)	
<b>E. SEPARATE FURNISHING OF INDICATIONS</b> (leave blank if not applicable)	
The indications listed below will be submitted to the International Bureau later (specify the general nature of the indications e.g., "Accession Number of Deposit")	

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## INDICATIONS RELATING TO A DEPOSITED MICROORGANISM

(PCT Rule 13bis)

A. The indications made below relate to the microorganism referred to in the description on page <u>6</u> , line <u>1</u>	
B. IDENTIFICATION OF DEPOSIT <span style="float: right;">Further deposits are identified on an additional sheet <input type="checkbox"/></span>	
Name of depositary institution AMERICAN TYPE CULTURE COLLECTION	
Address of depositary institution (including postal code and country) 12301 Parklawn Drive Rockville, Maryland 20852 US	
Date of deposit 12 June 1997	Accession Number 209120
C. ADDITIONAL INDICATIONS (leave blank if not applicable) <span style="float: right;">This information is continued on an additional sheet <input type="checkbox"/></span>	
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CLAIMS

1. An isolated nucleic acid fragment encoding a plant *p*-hydroxy-phenylpyruvate dioxygenase enzyme, the fragment comprising a nucleotide sequence selected from the group consisting of
  - 5 nucleotide sequences encoding a polypeptide comprising the amino acid sequences set forth in SEQ ID NO:3, SEQ ID NO:11, SEQ ID NO:13, and SEQ ID NO:15 and
  - modified nucleotide sequences essentially similar to the nucleotide sequences of SEQ ID NO:2, SEQ ID NO 10, SEQ ID NO:12 and
  - 10 SEQ ID NO:14 containing deletions, insertions, or substitutions in the sequence that do not affect the functional properties of the encoded protein.
2. An isolated nucleic acid fragment encoding a plant *p*-hydroxyphenylpyruvate dioxygenase enzyme, the fragment comprising a nucleotide sequence as set forth in SEQ ID NO:14.
- 15 3. A chimeric gene comprising the nucleic acid fragment of Claims 1 or 2 operably linked to at least one suitable regulatory sequence.
4. The chimeric gene of Claim 3 wherein at least one suitable regulatory sequence directs gene expression in a microorganism.
- 20 5. The chimeric gene of Claim 3 wherein the at least one suitable regulatory sequence directs gene expression in a plant.
6. A plasmid vector comprising the nucleic acid fragment of Claims 1 or 2 operablylinked to at least one suitable regulatory sequence.
7. A transformed host cell comprising a host cell and the plasmid vector
- 25 of Claim 6.
8. The transformed host cell of Claim 7 wherein the host cell is derived from a plant or is a microorganism.
9. The transformed host cell of Claim 8 wherein the microorganism is *E. coli*.
- 30 10. A transformed plant tolerant to contact with at least one compound that inhibits the rate of the reaction of *p*-hydroxyphenylpyruvate dioxygenase enzyme in a non-transformed plant, the transformed plant comprising the chimeric gene of Claim 3 and a host plant.
11. The transformed plant of Claim 10 wherein the host plant is a cereal crop plant.
- 35 12. A method to identify a compound useful for its ability to inhibit the rate of the reaction of *p*-hydroxyphenylpyruvate dioxygenase enzyme comprising:
  - (a) transforming a host cell with the plasmid vector of Claim 6;

- (b) facilitating expression of the nucleic acid fragment encoding the plant *p*-hydroxyphenylpyruvate dioxygenase enzyme;
- (c) contacting the expressed enzyme from step (b) with a test compound; and
- 5 (d) evaluating the capacity of the test compound to inhibit the rate of the reaction of *p*-hydroxyphenylpyruvate dioxygenase enzyme.

13. The method of Claim 12 wherein evaluating the capacity of the test compound to inhibit the rate of the reaction of *p*-hydroxyphenylpyruvate dioxygenase enzyme is accomplished by measuring oxygen utilization, carbon  
10 dioxide release, homogentisate production, loss of *p*-hydroxyphenylpyruvate or maleylacetoacetate production.

14. The method of Claim 12 wherein the transformed host cell is an *E. coli* that comprises a chimeric gene encoding a plant *p*-hydroxyphenylpyruvate dioxygenase enzyme.

15 15. A compound that inhibits the activity of a plant *p*-hydroxyphenylpyruvate dioxygenase enzyme, the compound identified by the method of Claim 14.

16. A method for imparting tolerance to a plant to at least one compound that inhibits the rate of reaction of *p*-hydroxyphenylpyruvate dioxygenase enzyme  
20 comprising:

- (a) transforming a host plant cell with a chimeric gene comprising a nucleic acid fragment encoding plant *p*-hydroxyphenylpyruvate dioxygenase, and
- (b) expressing the chimeric gene in an amount effective to render  
25 the transformed plant substantially tolerant to the at least one compound that inhibits the rate of reaction of *p*-hydroxyphenylpyruvate dioxygenase.

17. A method for the microbial production of active plant *p*-hydroxyphenylpyruvate dioxygenase enzyme comprising:

- 30 (a) stably transforming a microorganism with the chimeric gene of Claim 4 encoding the plant *p*-hydroxyphenylpyruvate dioxygenase;
- (b) facilitating expression by the chimeric gene for a suitable period; and
- 35 (c) recovering active plant *p*-hydroxyphenylpyruvate dioxygenase enzyme.

18. A method to overexpress *p*-hydroxyphenylpyruvate dioxygenase enzyme in a plant comprising:



- 5 (a) stably transforming a host plant cell with a chimeric DNA molecule comprising at least one copy of a suitable promoter to drive expression of an associated coding sequence in a plant cell operably linked to at least one copy of a homologous or heterologous coding sequence encoding *p*-hydroxyphenyl-pyruvate dioxygenase; and
- (b) growing the transformed host plant cell of step (a).
19. The method of Claim 18 wherein the chimeric DNA molecule is the chimeric gene of Claim 5.
- 10 20. An isolated nucleic acid fragment comprising a member selected from the group consisting of:
- 15 (a) an isolated nucleic acid fragment as set forth in SEQ ID NO:16;
- (b) an isolated nucleic acid fragment that is essentially similar to an isolated nucleic acid fragment as set forth in SEQ ID NO:16;
- and
- (c) an isolated nucleic acid fragment that is complementary to (a) or (b).

1/6

# FIG. 1

```
1   CAAGAAACGNGTCGNCGACGTGCTCAGCGATGATCAGATCAAGGAGTGTGAGGAATTAGG
61  GATTCTTNTAGACAGAGATGATCAAGGGACGTTNCTTCAAATCTNCACAAAACCACTAGG
121 TGACAGGCCGACGNTATTTATAGAGATAATCCAGAGNGTAGGATGCATGATGAAAGATGT
181 GGAAGGGANGGCTTACCAGAGTGGAGNATNTNGTGGTTTTGGCAAAGGCAATT
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2/6

## FIG. 2

1 TGAAATCAATGGGCCACCAAAACGCCGCCGTTTCAGAGAATCAAACCATGATGACGGCG  
61 CTGCGTCGTCGCCGGGATTCAAGCTCGTCGGATTTTCCAAGTTCGTAAGAAAGAATCCAA  
121 AGTCTGATAAATTCAAGGTTAAGCGCTTCCATCACATCGAGTCTGGTGCGGGGACGCAA  
Eco47III  
181 CCAACGTCGCTCGTCGCTTCTCCTGGGGTCTGGGGATGAGATTCTCGCCAAATCCGATC  
241 TTTCCACCGGAAACATGGTTCACGCCTCTTACCTACTCACCTCCGGTGAACCTCCGATTCC  
301 TTTTCACTGCTCCTTACTCTCCGTCTCTCTCCGGCGGAGAGATTAAACCGACAACCACAG  
361 GTTCTATCCCAAGTTTCGATCACGGGTCTTGTCGGTCCTTCTTCTTTCACATGGTCTCG  
421 GTGTTAGACCCGTTGCGATTGAAGTAGAAGACGCGGAGTCAGCTTTCTCCATCAGTGTAG  
481 CTAATGGCGCTATTCTTCGTCGCCTCTATCGTCCTCAATGAAGCAGTTACGATCGCTG  
541 AGGTTAAACTATACGGCGATGTTGTTCTCCGATATGTTAGTTACAAAGCAGAAGATACCG  
601 AAAAATCCGAATTCTTGCCAGGGTTCGAGCGTGTAGAGGATGCGTCGTCGTTCCCATTTGG  
EcoRI  
661 ATTATGGTATCCGGCGGCTTGACCACGCCGTGGGAAACGTTCTGAGCTTGGTCCGGCTT  
721 TAACTTATGTAGCGGGGTTCACTGGTTTTTACCAATTTCGCAGAGTTCACAGCAGACGACG  
781 TTGGAACCGCCGAGAGCGGTTTAAATTCAGCGGTCTTGGCTAGCAATGATGAAATGGTTC  
NheI  
841 TTCTACCGATTAAACGAGCCAGTGCACGGAACAAAGAGGAAGAGTCAGATTACAGACGTATT  
901 TGGAACATAACGAAGGCGCAGGGCTACAACATCTGGCTCTGATGAGTGAAGACATATTCA  
961 GGACCCTGAGAGAGATGAGGAAGAGGAGCAGTATTGGAGGATTCGACTTCATGCCTTCTC  
1021 CTCCGCCTACTTACTACCAGAATCTCAAGAAACGGGTCCGCGACGTGCTCAGCGATGATC  
1081 AGATCAAGGAGTGTGAGGAATTAGGGATTCTTGTTAGACAGAGATGATCAAGGGACGTTGC  
1141 TTCAAATCTTCACAAAACCACTAGGTGACAGGCCGACGATATTTATAGAGATAATCCAGA  
1201 GAGTAGGATGCATGATGAAAGATGAGGAAGGGAAGGCTTACCAGAGTGGAGGATGTGGTG  
1261 GTTTTGCCAAAGGCAATTTCTCTGAGCTCTTCAAGTCCATTGAAGAATACGAAAAGACTC  
1321 TTGAAGCCAAACAGTTAGTGGGATGAACAAGAAGAAGAACCAACTAAAGGATTGTGTAAT  
1381 TAATGTAAACTGTTTTATCTTATCAAAACAATGTATACAACATCTCATTTAAAAACGAG  
1441 ATCAATCC

3/6

## FIG. 3A

Arabidopsis	1	MGHQNAAVS	ENQNHDDGAA	SSPGFKLVGF	SKEVRKNPKS	DKFKVKRFHH	50
Corn		MPPTPTAAAA	GAAVAAASAA	EQAAFRVLGH	RNEVRFNPRS	CRFHTLAFHH	
Rat					YWDKGPKP	EPGRFLHFHS	
Mouse					M TTYNNKGPKP	ERGRFLHFHS	
Human					M TYSYDKGAKP	ERGRFLHFHS	
Pig					M TSYSDKGEKP	ERGRFLHFHS	**
Arabidopsis	51	IEFWCGDATN	VARRFSGWLG	MRFSAKSDLS	TGNMVHASYL	LTSGDLRFLF	100
Corn		VELWCADAAS	AAGRFSFGLG	APLAARSDLS	TGNSAHASLL	LRSGSLSFLL	
Rat		VTFWVGNAKQ	AASFYCNKMG	FEPLAYKGLE	TGSREVVSHV	IKQGKIVFVL	
Mouse		VTFWVGNAKQ	AASFYCNKMG	FEPLAYRGL	TGSREVVSHV	IKRGKIVFVL	
Human		VTFWVGNAKQ	AASFYCSKMG	FEPLAYRGL	TGSREVVSHV	IKQGKIVFVL	
Pig		VTFWVGNAKQ	AASYCSKIG	FEPLAYKGLE	TGSREVVSHV	VKQDKIVFVF	*
		*	*	*	*	*	*
Arabidopsis	101	TAPYSPSLSA	GEIKPTTAS	IPSFHDGSCR	SFFSSHGLGV	RAVAIEVEDA	150
Corn		TAPYAHGADA	.....ATAA	LPSFSAAAAR	RFAADHGLAV	RAVALRVADA	
Rat		CSALNPW...	.....	.....NKEMG	DHLVKHGDGV	KDIAFEVEDC	
Mouse		CSALNPW...	.....	.....NKEMG	DHLVKHGDGV	KDIAFEVEDC	
Human		SSALNPW...	.....	.....NKEMG	DHLVKHGDGV	KDIAFEVEDC	
Pig		SSALNPW...	.....	.....NKEMG	DHLVKHGDGV	KDIAFEVEDC	** * *
Arabidopsis	151	ESAFSISVAN	GAIPSSPPIV	LNEAVTIAEV	KLYGDVVLRY	VSYKAEDTEK	200
Corn		EDAFRASVAA	GARPAFGPVD	LGRGFRLAEV	ELYGDVVLRY	VSY. PDGAAG	
Rat		EHIVQKARER	GAKIVREPWW	EEDKFGKVKF	AVLQTYGDTT	HTLVEKINYT	
Mouse		DHIVQKARER	GAKIVREPWW	EODKFGKVKF	AVLQTYGDTT	HTLVEKINYT	
Human		DYIVQKARER	GAKIMREPWW	EODKFGKVKF	AVLQTYGDTT	HTLVEKMNYI	
Pig		DYIVQKARER	GAIIVREPWI	EODKFGKVKF	AVLQTFGDTT	HTLVEKMNYT	** *
Arabidopsis	201	SEFLPGFER.	..VEDASSFP	LDYGIRRLDH	AVGNVP..EL	GPALTYVAGF	250
Corn		EPFLPGFEG.	..V..ASPGA	ADYGLSRFDH	IVGNVP..EL	APAAAYFAGF	
Rat		GRFLPGFEAP	TYKDTLLPKL	PSCNLEIIDH	IVGNQPDQEM	ESASEWYLKN	
Mouse		GRFLPGFEAP	TYKDTLLPKL	PRCNLEIIDH	IVGNQPDQEM	QSASEWYLKN	
Human		GQFLPGYEPP	AFMDPLLPLK	PKCSLEMIDH	IVGNQPDQEM	VSASEWYLKN	
Pig		GCFLPGFEAP	TFTDPLLSKL	PKCGLEIIDH	IVGNQPDQEM	ESASQWYMRN	*
		**** *		** *	**** *		
Arabidopsis	251	TGFHQFAEFT	ADDVGTAESG	LNSAVLASND	EMVLLPINEP	VHGTRKRSQI	300
Corn		TGFHEFAEFT	TEDVGTAESG	LNSMVLANN	ENVLLPLNEP	VHGTRKRSQI	
Rat		LQFHREWSVD	DTQVHTEYSS	LRSIVVANYE	ESIKMPINEP	APG.RKKSQI	
Mouse		LQFHREWSVD	DTQVHTEYSS	LRSIVVTNIE	ESIKMPINEP	APG.RKKSQI	
Human		LQFHREWSVD	DTQVHTEYSS	LRSIVVANYE	ESIKMPINEP	APG.KKKSQI	
Pig		LQFHREWSVD	DTQIHTEYSA	LRSVVMANYE	ESIKMPINEP	APG.KKKSQI	***
		***	*	*	*	*	*

4/6

## FIG. 3B

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301                                     350
Arabidopsis QTYLEHNEGA GLQHLALMSE DIFRTLREMR KRSSIGGFDF MPSPPTYYQ
  Corn QTFLDHHGGP GVQHMALASD DVLRTLREMQ ARSAMGGFEF MAPPTSDDYYD
  Rat QEYVDYNGGA GVQHIALRTE DIITTIRHLR ER....GMEF LAVP.SSYYR
  Mouse QEYVDYNGGA GVQHIALKTE DIITAIRHLR ER....GTEF LAAP.SSYYK
  Human QEYVDYNGGA GVQHIALKTE DIITAIRHLR ER....GLEF LSVP.STYYK
  Pig QEYVDYNGGA GVQHIALKTE DIITAIRSLR ER....GVEF LAVP.FTYYK

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351                                     400
Arabidopsis NLKK..RVGD VLSDDQIKEC EELGILVDRD DQGTLLQIFT KPLGDRPTIF
  Corn GVRR..RAGD VLTEAQIKEC QELGVLVDRD DQGVLLQIFT KPVGDRPTLF
  Rat LLRENLKTSK IQVKENMDVL EELKILVDYD EKGYLELQIFT KPMQDRPTLF
  Mouse LLRENLKSAK IQVKESMDVL EELHILVDYD EKGYLELQIFT KPMQDRPTLF
  Human QLREKLKTAK IKVKENIDAL EELKILVDYD EKGYLELQIFT KPVQDRPTLF
  Pig QLQEKLSAK IRVKESIDVL EELKILVDYD EKGYLELQIFT KPMQDRPTVF

```

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401                                     450
Arabidopsis IEIIQRVGCM MKDEEGKAYQ SGGCGGFGKG NFSELFKSIE EYEKTLQAKQ
  Corn LEIIQIRIGCM EKDEKGQEQY KGGCGGFGKG NFSOLFKSIE DYEKSLEAKQ
  Rat LEVIQRHNNHQ .....GFGAG NFNSLFKAFE E.EQALRG
  Mouse LEVIQRHNNHQ .....GFGAG NFNSLFKAFE E.EQALRGNL
  Human LEVIQRHNNHQ .....GFGAG NFNSLFKAFE E.EQNLRGNL
  Pig LEVIQRNNHQ .....GFGAG NFNSLFKAFE E.EQELRGNL

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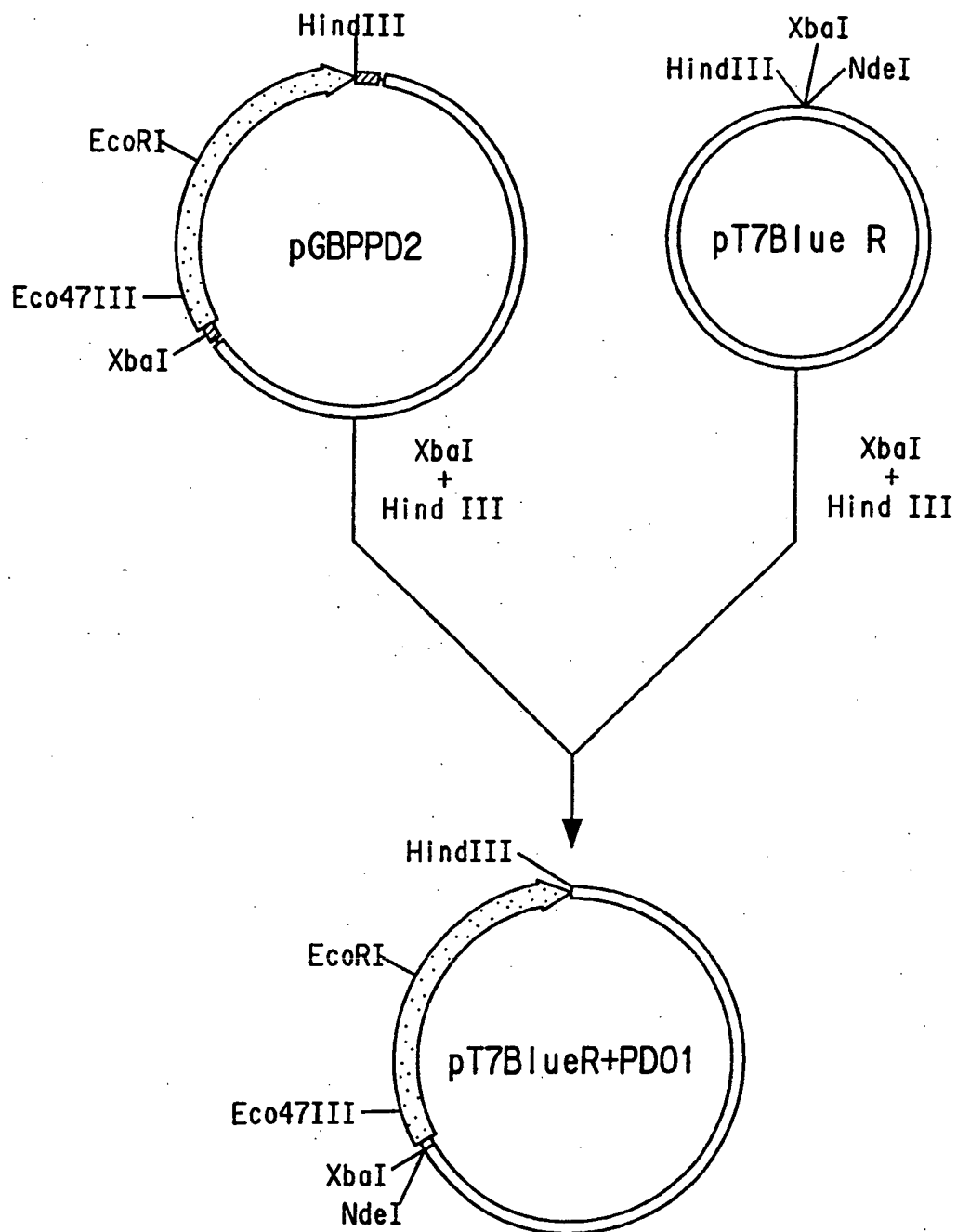
```

451         462
Arabidopsis LVG (Seq. I.D. No. 15)
  Corn AAAAAAAGGS (Seq. I.D. No. 11)
  Rat (Seq. I.D. No. 9)
  Mouse TDLEPNGVRS GM (Seq. I.D. No. 8)
  Human TNMETNGVVP GM (Seq. I.D. No. 6)
  Pig TDTDPNGVPP RL (Seq. I.D. No. 7)

```

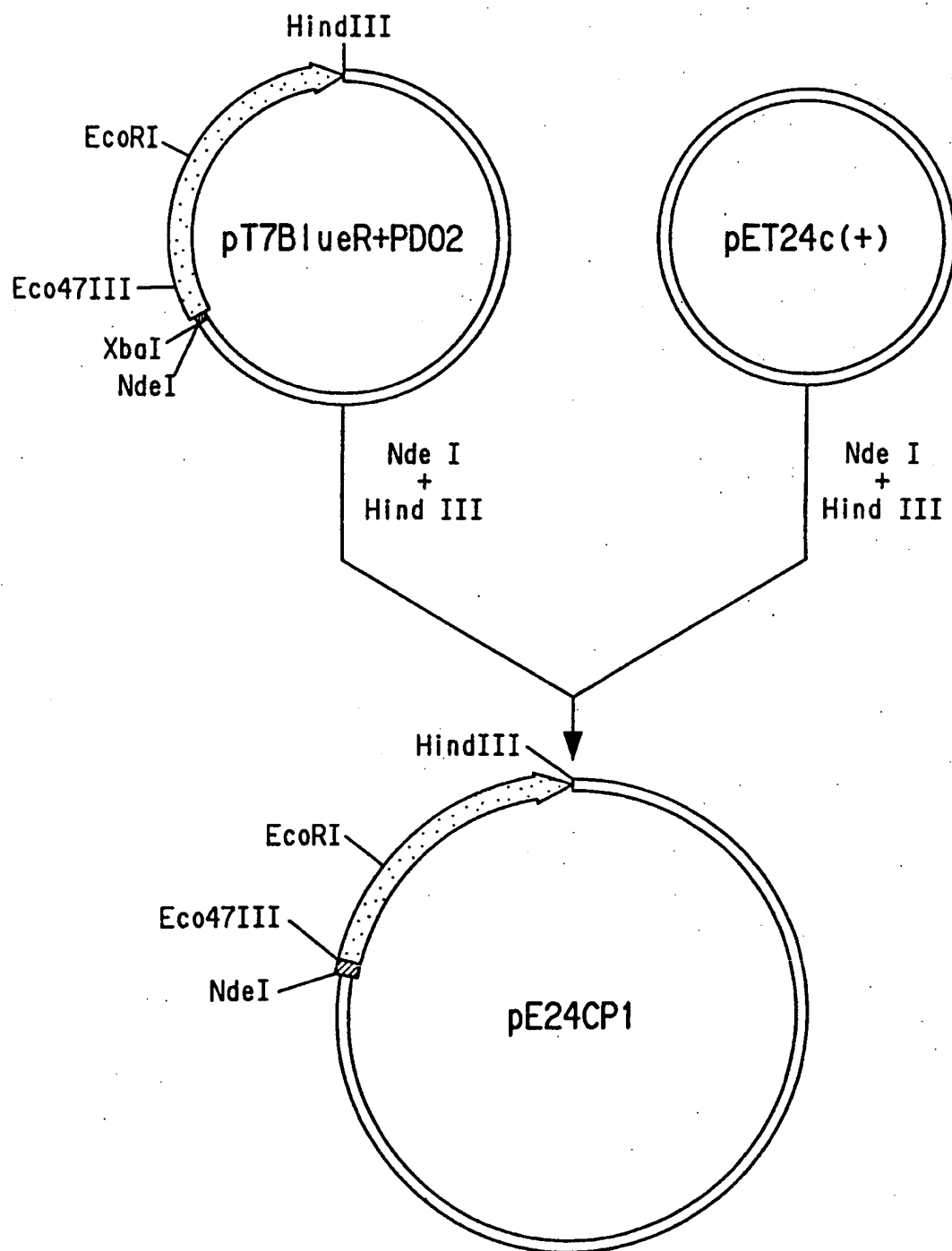
5/6

FIG. 4



6/6

FIG. 5



# INTERNATIONAL SEARCH REPORT

International Application No  
PCT/US 97/11295

A. CLASSIFICATION OF SUBJECT MATTER  
IPC 6 C12N15/53 C12N15/82 C12Q1/26 C12Q1/02 A01H5/00

According to International Patent Classification (IPC) or to both national classification and IPC

## B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

IPC 6 C12N C12Q A01H

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

## C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No. .
X	NEWMAN, T., ET AL.: "2960 Arabidopsis thaliana cDNA clone 91B13T7" EMBL SEQUENCE DATABASE, REL. 40, 16-JUN-1994, ACCESSION NO. T20952, XP002028637 see sequence	1,2
X	NEWMAN, T., ET AL.: "20804 Arabidopsis thaliana cDNA clone 231K20T7" EMBL SEQUENCE DATABASE, REL.47, 8-MAR-1996, ACCESSION NO. N65764, XP002029449 see sequence	1,2



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Date of the actual completion of the international search

26 September 1997

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## C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
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A	EP 0 652 286 A (RHONE POULENC AGROCHIMIE) 10 May 1995 see page 7, line 35 - line 47 ---	10,16,18
A	MISAWA N ET AL: "EXPRESSION OF AN ERWINA PHYTOENE DESATURASE GENE NOT ONLY CONFERS MULTIPLE RESISTANCE TO HERBICIDES INTERFERING WITH CAROTENOID BIOSYNTHESIS BUT ALSO ALTERS XANTHOPHYLL METABOLISM IN TRANSGENIC PLANTS" PLANT JOURNAL, vol. 6, no. 4, 1994, pages 481-489, XP002017203 see the whole document ---	10,16,18

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# INTERNATIONAL SEARCH REPORT

International Application No.

PCT/US 97/11295

## C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
A	DENOYA C D ET AL: "A STREPTOMYCES AVERMITILIS GENE ENCODING A 4-HYDROXYPHENYLPYRUVIC ACID DIOXYGENASE-LIKE PROTEIN THAT DIRECTS THE PRODUCTION OF HOMOGENTISIC ACID AND AN OCHRONOTIC PIGMENT IN ESCHERICHIA COLI" JOURNAL OF BACTERIOLOGY, vol. 176, no. 17, September 1994, pages 5312-5319, XP002028042 see the whole document	17
A	NORRIS, S.R., ET AL.: "Genetic dissection of carotenoid synthesis in Arabidopsis defines plastoquinone as an essential component of phytoene desaturation" THE PLANT CELL, vol. 7, December 1995, pages 2139-2149, XP002041909 cited in the application see the whole document	1-20

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Information on patent family members

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PCT/US 97/11295

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